

Review Article

Association of Peroxisome Proliferator-Activated Receptors (PPARs) with Diabetic Retinopathy in Human and Animal Models: Analysis of the Literature and Genome Browsers

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Diabetic retinopathy (DR) is a condition that develops after long-lasting and poorly handled diabetes and is presently the main reason for blindness among elderly and youth. Peroxisome proliferator-activated receptors (PPARs) are nuclear receptors that are involved in carbohydrate and fatty-acid metabolism and have also been associated with DR. Three PPAR isoforms are known: PPARG, PPARA, and PPARD. In the present study, we retrieved articles reporting associations between PPARs and DR from PubMed database and compiled the data in two catalogues, for human and animal models. Extracted data was then complemented with additional relevant genomic information. Seven retrieved articles reported testing an association between PPARs with DR in human. Four of them concluded association of PPARG and PPARA with DR in European and Asian populations, having a protective role on DR development. One study reported pathogenic role of PPARG, while two articles reported no association between PPARG and DR among Indian and Chinese populations. Six retrieved articles reported testing of involvement of PPARG and PPARA in DR in animal models, including mouse and rat. The review includes case-control studies, meta-analysis, expression studies, animal models, and cell line studies. Despite a large number of documented sequence variants of the PPAR genes available in genome browsers, researchers usually focus on a small set of previously reported variants. Data extraction from Ensembl genome browser revealed several sequence variants with predicted deleterious effect on protein function which present candidates for further experimental validation. Results of the present analysis will enable more holistic approach for understanding of PPARs in DR development. Additionally, developed catalogues present a baseline for standardized reporting of PPAR-phenotype association in upcoming studies.

1. Introduction

Diabetic retinopathy (DR) is a condition that develops due to bad glycemic control in subjects with type 1 diabetes mellitus (T1DM) or type 2 diabetes mellitus (T2DM). Long-lasting poor blood glucose control, smoking, and hypertension can contribute to DR development [1, 2]. The disease progresses from nonproliferative (NPDR) to proliferative (PDR) stage where at first microvascular irregularities such as hemorrhage, ischemia, and microaneurysms lead to neoangiogenesis [2]. Microvascular changes start due to lower concentrations of oxygen in the retina of the eye after the disease progresses, and at final stages, PDR can lead to vision loss. Diabetic retinopathy had become the main reason for blindness in American adults. In year 2012, there were approximately 93 million people living with diabetic retinopathy, 17 million with PDR, and 21 with diabetic macular edema, and the number is expected to increase in the future [3, 4].

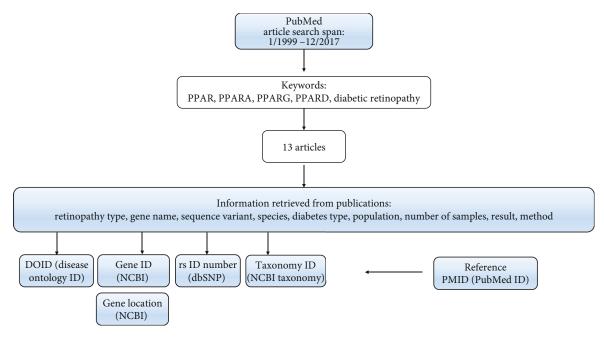


FIGURE 1: Workflow of the study.

Peroxisome proliferator-activated receptors (PPARs) are nuclear receptors that regulate the expression of several genes and are affecting lipid and carbohydrate metabolism. PPARs consist of three subtypes: PPARA, PPARD, and PPARG [5]. Peroxisome proliferator-activated receptor gamma (PPARG) also known as GLM1, CIMT1, NR1C3, PPARG1, PPARG2, or PPARgamma is a nuclear receptor that binds hypolipidemic drugs and unsaturated fatty acids and affects adipocyte differentiation, gluconeogenesis, oxidation of fatty acids, lipogenesis, cholesterol metabolism, and synthesis of ketone bodies [5-9]. The gene is located on chromosome HSA3. In the eye, the gene is heterogeneously expressed in photoreceptor outer segments, choriocapillaries, retina, retinal pigmented epithelium, cornea, and lacrimal gland [10-12]. Three RNA isoforms of expressed PPARG have been identified: y1, y2, and γ 3. PPAR- γ 2 protein has additional stretch of 28 amino acids on N-terminal, and this extension seems to change PPAR-y2 sensitivity to insulin action [13]. Proline variant of Pro12Ala (rs1801282) polymorphism of the PPARG gene is associated with increased resistance to insulin action whereas the alternative allele has the opposite properties [14].

Peroxisome proliferator-activated receptor alpha (*PPARA*) also known as *PPARα*, *NR1C1*, *hPPAR*, or *PPARalpha* is responsible for ketogenesis, lipid transport, lipogenesis, cholesterol metabolism, fatty acid transport, and oxidation [15]. It is located on the HSA22. *PPARA* is expressed in the retina; however, its levels have been shown to be reduced in the retinas with DR [16, 17]. Decreased *PPARA* expression in diabetic retinas contributes to retinal inflammation and neovascularization in DR, and activation of PPARA has anti-inflammatory and antiapoptotic effects in oxygen-induced retinopathy (OIR) and diabetic animal models through suppression of *NF-κB* signaling [16, 17].

Peroxisome proliferator-activated receptor delta (PPARD) also known as *FAAR*, *NUC1*, *NUCI*, *NR1C2*, *NUCII*, or *PPARB* is located on HSA6. It affects fatty acid transport and

oxidation, adipocyte differentiation, adaptive thermogenesis, cell survival, and ubiquitination [18]. Among the three PPAR subtypes, it is the least studied and understood, especially its effects on inflammation and proliferation associating DR.

To our knowledge, the complete database related with reported associations between PPARs and DR does not yet exist. The aim of this study was therefore to conduct an overview of articles reporting an association between three PPARs and DR/PDR in human and animal models.

2. Materials and Methods

Using keywords "PPAR" and/or "PPARG" and/or "PPARA" and/or "PPARD" and/or "polymorphism" and/or "diabetic retinopathy", we explored the PubMed database for articles describing association between PPARs and DR in human and animal models. Inclusion criteria for the type of study in humans were case-control study, meta-analysis, or expression study. Retrieved articles included in previously published meta-analysis were excluded from the analysis. Time span for article search was set from January 1999 to December 2017. Retrieved articles were checked for the following information: retinopathy type, sequence variant, gene name, diabetes type, species, number of tested samples, result of the study, and method. The data extracted from publications was afterwards complemented with additional information such as gene ID (https://www.ncbi.nlm.nih.gov/ gene), gene location (https://www.ncbi.nlm.nih.gov/gene), taxonomy ID (https://www.ncbi.nlm.nih.gov/taxonomy), disease ontology ID (DOID; http://disease-ontology.org/), reference SNP (rs) identification number, PubMed identification number (PMID) of the reference, and statistical significance (Figure 1). Ensembl genome browser release 96 was used to retrieve additional information related with sequence variants, predicted effect on protein function

using six bioinformatics tools, and clinical significance from ClinVar database [19].

3. Results

We developed two tables consisting of data extracted from 13 retrieved articles published between 1/2012 and 12/2017 reporting associations between PPAR polymorphisms and DR in human (Table 1) and animal models (Table 2) (Figure 2). In humans, six articles reported testing association between *PPARG* and DR/PDR and one reported *PPARA* and DR association. We did not retrieve any articles related with *PPARD*-DR association. Six studies were performed in animal models, including four articles describing involvement of *PPARA* in DR and two involvement of *PPARG* in DR.

3.1. Studies in Humans. Out of seven retrieved articles describing association between *PPARs* and DR/PDR in humans, six articles were related with the *PPARG* gene and one study with the *PPARA* gene.

One study reported that *PPARG* may play an important role in the pathogenesis of PDR. The *PPARG* concentrations in the aqueous humor and vitreous fluid were significantly higher in PDR patients than in controls, and the level of *PPARG* increased in the advanced clinical stage. Additionally, a correlation between *PPARG* and vascular endothelial growth factor (*VEGF*) concentrations was identified [20]. Two out of seven studies reported no association between *PPARs* and DR [21, 22]. Three studies identified association (protective effect or decreased DR risk) between *PPARG* and DR/PDR (Figure 2) [23–25]. Qi et al. studied polymorphism rs1800206 of the *PPARA* gene and concluded that carriers of homozygous mutant allele have decreased DR risk in comparison to wild-type homozygotes in Chinese Han population [26].

Most participants in the studies had type 2 diabetes mellitus (T2DM), and some participants had type 1 diabetes mellitus (T1DM). The developed catalogue includes five case-control studies, one meta-analysis [23], and one expression study [20]. Case-control studies included 17 to 812 participants. Meta-analysis study consisted of more than 4000 participants from eight studies. Studies were performed on different populations, such as European Caucasian, Asian (Chinese Han), and Pakistani. Methods used for genotyping and expression analysis were quantitative real time, PCRligase detection reaction (LDR), quantitative PCR, PCR-RFLP, and real-time PCR.

3.2. Studies in Animal Models. Six studies used animal models for testing association between *PPARs* and DR/PDR: mouse, rat, and cattle. In some studies, more than one animal model and additional animal cell lines were used. For imitating DR or diabetes in mice and rat, animals were made diabetic with streptozotocin (STZ) or underwent through OIR. Most studies based on an animal model used knockout mice (KO) approach. In most studies, they used C57Bl/6J mouse model or Brown Norway rats [16, 17, 27–30]. Additionally, bovine retinal endothelial cells (BRECs) were also used [28].

Various methods were used for testing association between PPARs and DR in animal models, for example, TUNEL assay, quantitative real-time PCR, retinal leakage assay, vascular leakage assay, fluorescent microscopy, immunofluorescence, western blot, and protein-based detection methods detecting over/underexpression of the protein.

Most of the reports in humans were designed as association studies between *PPAR* polymorphisms and DR; however, in animal models, most performed gene expression analyses in diabetic and nondiabetic animals (Table 2). Hu et al. [17] used animal model for testing an involvement of *PPARA* and DR and concluded that *PPARA* knockout mice developed more severe DR which resulted in retinal vascular leakage, leukostasis, pericyte loss, capillary degeneration, and overexpressed inflammatory factors, whereas *PPARA* overexpression reduced vascular leakage and inflammation. *PPARA* protective effects have been proven by Ding et al. [30]. *PPARG*+/- knockout mice had greater leukostasis and leakage than wild-type mice [27], and suppression of *PPARG* has been shown to be involved in the pathogenesis of diabetic retinopathy and OIR [28].

4. Discussion

PPARs are important factors in DR/PDR due to their protective function on the disease development. Our results revealed that reports in this study field are very heterogeneous. Most studies in humans analyzed polymorphism Pro12Ala (rs1801282) located in the *PPARG* gene. In contrary, some studies were performed on cell lines and animal models. For example, Chen et al. [29] reported that *PPARA* is a target of microRNA-21, which downregulates expression of *PPARA* and worsens DR condition.

Our study revealed that researchers use different synonyms for the same gene (for example, *PPARG*, *PPARγ*, *CIMT1*, and *NR1C3*), for the same gene variant (Pro12Ala, rs1801282, c.34C>G), or for methodology. In several studies, patients with DR were not divided into NPDR and PDR cases. Additionally, in some studies, it is not clear whether a gene is associated with PDR or is associated only with NPDR.

The results of the association studies related with PPARs and its association with DR/PDR differ among populations (Table 1). For example, polymorphism Pro12Ala is the most studied polymorphism of the *PPARG* gene. Tariq et al. reported that polymorphism Pro12Ala is not associated with DR in Pakistani population [25]; however, Wang et al. reported that it is associated with DR in Chinese population [24].

According to the latest release of the Ensembl database, there are a high number of polymorphisms located within *PPAR* genes in humans and animals. However, our results show that researchers focused on only few sequence variants of the *PPAR* gene family. Several bioinformatics tools could be used for prioritization of stronger candidate sequence variants for experimental validation. Ensembl browser enables comparison of six bioinformatics tools designed for predicting the effect of sequence variants on protein function: SIFT, PolyPhen, CADD. REVEL, MetaLR, and MutationAssessor. Figure 3 presents a part of the variant table from the Ensembl genome browser. For example, most of the tools predict

PMID	25468312	25274455	22993484	26885119	23559865	27427939	26671228
Reference	Katome et al. [20]	Zhang et al. [22]	Ma et al. [23]	Wang et al. [24]	Tariq et al. [25]	Kaur et al. [21]	Qi et al. [26]
Type of study	Expression study	Case- control study	Meta- analysis	Case- control study	Case- control study	Case- control study	Case- control study
Main result of the study	Higher expression of <i>PPARG</i> in PDR versus controls	No significant association between polymorphisms in the <i>PPARG</i> gene and DR or PDR	Protective effect of Pro12Ala on DR in T2DM with ethnic differences	rs1805192 minor allele (Ala) of <i>PPARG</i> is significantly associated with lower DR risk; combined effect of Ala-BMI interaction between polymorphism and overweight on DR	Protective role of the 12Ala polymorphism against PDR in T2DM	No significant association	Association between rs1800206 minor (Y) allele and lower risk for DR; interaction between rs1800206 and abdominal obesity
Method	Quantitative real-time PCR, ELISA, immunohisto- chemistry analysis	PCR-LDR	Statistics	Quantitative PCR	PCR-RFLP	Real-time PCR	Quantitative PCR
Statistical significance	<i>p</i> < 0.0005	OR (95% CI) dominant model GG: 1.40 (0.85-2.29); $p = 0.22$	Caucasian subgroup (OR = $0.74;$ 95% CI: $0.59-$ 0.94, p = 0.01) Asian subgroup (OR = $0.7; 95\%$ CI: $0.55-1.07;$ p = 0.12)	OR (95%CI) = 0.86 (0.65-0.96), <i>p</i> = 0.012	OR = 0.4; 95%CI = 0.2-0.8	<i>p</i> = 0.507	OR (95%CI) = 0.78 (0.66-0.94)
Number of samples (cases/controls)	17 (12 PDR, 5 controls)	792 T2DM (448 DR, 344 diabetes without DR)	5170 (2720 DR cases, 2450 controls)	500 T2DM (247 DR cases, 253 controls)	573 (189 DR, 193 DNR, 200 controls)	1325 (717 DR, 608 T2DM without DR)	812 (402 DR, 410 control)
Population	Japan*	Chinese	Caucasian Asian	Chinese	Pakistani	Indian	Chinese Han
DOID	13207	8947, 13207	8947	8947	8947, 13207	8947	8947
Retinopathy type	PDR	DR, PDR	DR	DR	DR, PDR	DR	DR
Diabetes type	T1DM T2DM	T2DM	T2DM	T2DM	T2DM	T2DM	T2DM
rs ID of the polymorphism	~	rs1801282 rs3856806 rs12497191	~	rs3856806 rs709158 rs1805192 rs4684847	rs1801282	-	rs4253778 rs135539 rs1800206
Sequence variant		rs1801282 rs3856806 rs12497191	Pro12Ala	C1341T Intron A>C Pro12Ala Intron C>T	rs1801282 (c.34C>G, Pro12Ala)	p.Pro12Ala	rs4253778 rs135539 rs1800206
Gene location	3p25.2	3p25.2	3p25.2	3p25.2	3p25.2	3p25.2	22q13.31
Gene ID	5468	5468	5468	5468	5468	5468	5465
Gene symbol	PPARG	PPARG	PPARG	PPARG	PPARG	PPARG	PPARA

4

PMID	17003451	17003451	18806296	18806296	28270521	28979999	28979999	25108226	24003152	24003152
Reference	Muranaka et al. [27]	Muranaka et al. [27]	Tawfik et al. [28]	Tawfik et al. [28]	Chen et al. [29]	Deng et al. [16]	Deng et al. [16]	Ding et al. [30]	Hu et al. [17]	Hu et al. [17]
Type of study	Animal model	Animal model	Animal model	Animal model	Cell line, animal model	Cell line, animal model	Cell line, animal model	Animal model	Animal model	Animal model
Main result of the study	<i>Pparg</i> signaling pathway inhibits diabetes-induced retinal leukostasis and leakage	Therapy with <i>Pparg</i> ligands may inhibit retinal leukostasis and retinal leakage in diabetes	The link between <i>Pparg</i> and retinal vascular inflammation in DR	Suppression of <i>Pparg</i> expression in high glucose-treated cells	Upregulated <i>miR-21</i> and downregulated <i>Ppara</i> in OIR	Y-0452 exerts antiangiogenic effects in OIR retinas through <i>Ppara</i> -dependent mechanism	Y-0452 (<i>Ppara</i> agonist) alleviated the retinal apoptosis	Protective effect of <i>Ppara</i> against retinal pericyte loss in DR	<i>Ppara</i> knockout mice developed more severe DR	Overexpression of <i>Ppara</i> in the retina alleviated vascular leakage and inflammation
Method	Retinal leakage assay, fluorescent microscopy	Retinal leakage assay, fluorescent microscopy	Immunofluo- rescence, western blot	Western blot	qRT-PCR	TUNEL assay	Vascular leakage assay	TUNEL assay	Quantitative real-time RT-PCR	Quantitative real-time RT-PCR
Statistical significance	<i>p</i> < 0.05	<i>p</i> < 0.05	<i>p</i> < 0.05	<i>p</i> < 0.05	<i>p</i> < 0.05	$p \leq 0.05$	$p \leq 0.05$	<i>p</i> < 0.05	<i>p</i> < 0.05	<i>p</i> < 0.05
Strain/details	C57BL/6 PPARg(+/-)	Brown Norway	C57BI/6J	BRECs	C57BLKS/J C57BL/6J	Ppara(-/-) C57/BL6J	Brown Norway	Ppara(-/-) C57/BL6J	C57BL/6J PPARA knockout Akita db/db	Brown Norway
DOID	8947	8947	8947	8947	8947	8947	8947	8947	8947	8947
Ketinopathy type model	DR	DR	DR	DR	DR	DR	DR	DR	DR	DR
Model	Knockout, STZ	STZ	Knockout, STZ, OIR	Cells	Knockout, OIR, cells	Knockout, OIR	STZ	Knockout, STZ	Knockout, STZ	STZ
Sequence variant	~	~	~	~	~	~	~	~	~	~
Taxonomy ID	10090	10116	10090	9913	10090	10090	10116	10090	10090	10116
Species	Mouse	Rat	Mouse	Cattle	Mouse	Mouse	Rat	Mouse	Mouse	Rat
location	6 E3	4q42	6 E3	22q24	15 E2	15 E2	7q34	15 E2	15 E2	7q34
ID	19016	25664	19016	281993	19013	19013	25747	19013	19013	25747
Gene symbol	Pparg	Pparg	Pparg	PPARG	Ppara	Ppara	Ppara	Ppara	Ppara	Ppara

PPAR Research

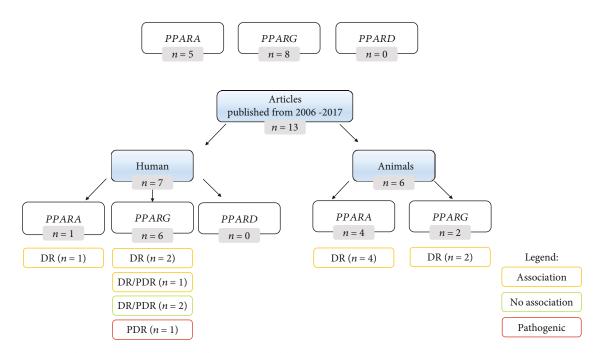


FIGURE 2: Overview of the results. The number of retrieved articles associating PPARs with DR/PDR is marked in grey color. Yellow color represents the number of articles reporting gene-DR/PDR association including protective effect or decreased DR risk. Green color represents the number of articles reporting no association between the gene and DR/PDR, and red color represents the number of articles reporting a pathogenic effect on DR/PDR.

Variant ID	Chr: bp	Alle- les	Glo- bal MAF	Class	Source	Evidence	Clin. Sig.	Conseq. Type	AA	AA co- ord		Poly- Phen	CAD D	REV EL	Meta LR	Mutat ion Asse ssor
<u>rs1801282</u>	3:12351626	C/G	0.070 (G)	SNP	dbSNP		*	missense variant	P/A	12	0.05	0) 17	0.238	0.019	0.18
<u>rs775382056</u>	3:12392651	A/G	-	SNP	dbSNP		-	missense variant	Y/C	173	0	1	29	0.98	0.959	0.91
<u>rs971658739</u>	3:12392700	A/T	ē.,	SNP	dbSNP	5	7	missense variant	K/N	189	0	1	25	0.774	0.929	0.418
rs121909245	3:12392701	T/A		SNP	dbSNP	5	▲	missense variant	C/S	190	0	0.999	28	0.978	0.996	0.99
rs121909246	3:12392713	C/T	-	SNP	dbSNP	snom AD	A	missense variant	R/W	194	0	1) 32	0.957	0.983	0.99
<u>rs148195788</u>	3:12392714	G/A	-	SNP	dbSNP	🔮 📮 🕑 🍡	A	missense variant	R/Q	194	0	0.999	27	0.894	0.979	0.977
<u>rs1462603839</u>	3:12392734	G/A	-	SNP	dbSNP	AD gnom	÷	missense variant	V/M	201	0	1	33	0.828	0.943	0.514

FIGURE 3: Print screen of the Ensembl genome browser presenting a part of the variant table of the PPARG gene. The table includes seven selected sequence variants and results of bioinformatics prediction of their effect on protein function. Ensembl browser includes six tools for predicting effects of substitutions on protein function: SIFT, PolyPhen, CADD. REVEL, MetaLR, and Mutation Assessor. Predicted effect on protein function is shown in different colors: benign: green; tolerated/neutral: blue; possibly damaging: orange; damaging: red. Clin. Sig.: clinical significance; a classification of a variant's impact on disease, according to the ClinVar database: pathogenic: red triangle; likely pathogenic: orange triangle; likely benign: blue cross.

benign effect of the polymorphism rs1801282 (Pro12Ala) on protein function and two predict tolerated/neutral effect. On the contrary, for several other polymorphisms, predicted effect on protein function is damaging (red color) or possibly damaging (orange). Out of 286 sequences with available bioinformatics predictions, only polymorphism rs121909246 has predicted deleterious effect by all six bioinformatics tools. Additionally, according to the ClinVar database, this polymorphism has a pathogenic effect. However, several other missense polymorphisms of the *PPARG* gene have not yet been tested for association with diseases, including DR. For some of the polymorphisms, minor allele frequency (MAF) and clinical significance from ClinVar database are given. Currently, the Ensembl browser lists 10 sequence variants of the *PPARG* gene with pathogenic clinical effect extracted from the ClinVar database. Further bioinformatics prioritization studies should be performed for all three *PPAR* genes to select novel candidate loci for functional analyses. However, it should be noted that bioinformatics predictions obtained using these tools often differ and are not always consistent with results of experimental validation.

Several other approaches were also used for testing involvement of PPAR in DR and development of novel therapies [16, 31]. Dou et al. analyzed substances targeting PPARs in association with DR/PDR [32]. Human cell lines were used as model for testing the association between gene and disease [17, 30, 33]. Many drugs such as fibrates and thiazolidinediones have been widely used for treatment of dyslipidemia and have also been found to have direct association with PPARs. Fibrates are amphipathic carboxylic acids, and its therapeutic effects are PPARA dependent, which makes fibrates selective agonists of PPARA. Chen et al. [34] provided evidence of association between fibrates and PPARA with different PPARA agonists, PPARA antagonists, and PPARA-/- knockout mice and stated that fenofibrates work as PPARA activators which leads to transcription activation or inhibition of PPARA target genes which further leads to slower DR progression. Rosiglitazone, an antidiabetic drug from thiazolidinedione class, works as a PPARG agonist, and its work of action has been studied in diabetic mice [27].

Besides members of the *PPAR* family, several other genes have been tested in association with DR development, for example, *PPARGC1* and *VEGF* [35, 36]. Therefore, catalogues developed in the present study should be extended with additional genes, tested for association with DR. For future publications, it would be suggested to present the data with an official gene name and with rs ID name of the polymorphism. It is also suggested that researchers clearly state if the gene is associated with DR, PDR, or NPDR. Standardized reporting of genotype-phenotype is important for further research on this prominent topic for easier overview on the data to make new associations on PPARs and DR development and finding new targets for DR treatment.

5. Conclusions

PPARs are important protective factors of DR/PDR among certain populations and have potential for therapeutic targets. To the best of our knowledge, this is the first overview on the topic on PPARs associated with DR/PDR in human and animal models. The study presents a baseline for further studies, for example, meta-analyses and bioinformatics prioritization of new candidates for functional studies.

Additional Points

Executive Summary. (i) Literature review of studies testing an association between PPARs and diabetic retinopathy in human and animal models. (ii) Results showed that published results are opposing and data presentation of results in publications is heterogeneous. (iii) Developed catalogues summarizing PPAR-DR associations present a baseline for

standardized reporting of PPAR-phenotype association in upcoming studies. (iv) Prioritization of novel candidate sequence variants for further experimental validation using six bioinformatics tools revealed several substitutions with predicted deleterious effect on protein function.

Conflicts of Interest

We declare that there are no conflicts of interests.

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References

- G. J. Molla, F. Ismail-Beigi, B. Larijani et al., "Smoking and diabetes control in adults with type 1 and type 2 diabetes: a nationwide study from the 2018 National Program for Prevention and Control of Diabetes of Iran," *Canadian Journal of Diabetes*, 2019.
- [2] G. Liew, R. Klein, and T. Y. Wong, "The role of genetics in susceptibility to diabetic retinopathy," *International Ophthalmol*ogy Clinics, vol. 49, no. 2, pp. 35–52, 2009.
- [3] J. W. Y. Yau, S. L. Rogers, R. Kawasaki et al., "Global prevalence and major risk factors of diabetic retinopathy," *Diabetes Care*, vol. 35, no. 3, pp. 556–564, 2012.
- [4] R. Lee, T. Y. Wong, and C. Sabanayagam, "Epidemiology of diabetic retinopathy, diabetic macular edema and related vision loss," *Eye and Vision*, vol. 2, no. 1, p. 17, 2015.
- [5] The UniProt Consortium, "UniProt: a worldwide hub of protein knowledge," *Nucleic Acids Research*, vol. 47, no. D1, pp. D506–D515, 2019.
- [6] I. Issemann and S. Green, "Activation of a member of the steroid hormone receptor superfamily by peroxisome proliferators," *Nature*, vol. 347, no. 6294, pp. 645–650, 1990.
- [7] B. N. Finck, "The PPAR regulatory system in cardiac physiology and disease," *Cardiovascular Research*, vol. 73, no. 2, pp. 269–277, 2007.
- [8] S. Z. Duan, M. G. Usher, and R. M. Mortensen, "PPARs: the vasculature, inflammation and hypertension," *Current Opinion in Nephrology and Hypertension*, vol. 18, no. 2, pp. 128– 133, 2009.
- [9] Y.-X. Wang, "PPARs: diverse regulators in energy metabolism and metabolic diseases," *Cell Research*, vol. 20, no. 2, pp. 124– 137, 2010.
- [10] A. A. Herzlich, J. Tuo, and C.-C. Chan, "Peroxisome proliferator-activated receptor and age-related macular degeneration," *PPAR Research*, vol. 2008, Article ID 389507, 11 pages, 2008.
- [11] C. Beauregard and P. C. Brandt, "Peroxisome proliferatoractivated receptor agonists inhibit interleukin-1betamediated nitric oxide production in cultured lacrimal gland acinar cells," *Journal of Ocular Pharmacology and Therapeutics*, vol. 19, no. 6, pp. 579–587, 2003.
- [12] M. A. Sarayba, L. Li, T. Tungsiripat et al., "Inhibition of corneal neovascularization by a peroxisome proliferator- activated receptor- γ ligand," *Experimental Eye Research*, vol. 80, no. 3, pp. 435–442, 2005.

- [13] C.-J. Yen, B. A. Beamer, C. Negri et al., "Molecular scanning of the human peroxisome proliferator activated receptor γ (hPPARγ) gene in diabetic Caucasians: identification of a Pro12Ala PPARγ2 missense mutation," *Biochemical and Biophysical Research Communications*, vol. 241, no. 2, pp. 270– 274, 1997.
- [14] K. Hara, T. Okada, K. Tobe et al., "The Pro12Ala Polymorphism in PPARγ2 May Confer Resistance to Type 2 Diabetes," *Biochemical and Biophysical Research Communications*, vol. 271, no. 1, pp. 212–216, 2000.
- [15] T. C. Leone, C. J. Weinheimer, and D. P. Kelly, "A critical role for the peroxisome proliferator-activated receptor α (PPARα) in the cellular fasting response: the PPARα-null mouse as a model of fatty acid oxidation disorders," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 13, pp. 7473–7478, 1999.
- [16] G. Deng, E. P. Moran, R. Cheng et al., "Therapeutic effects of a novel agonist of peroxisome proliferator-activated receptor alpha for the treatment of diabetic retinopathy," *Investigative Opthalmology & Visual Science*, vol. 58, no. 12, pp. 5030– 5042, 2017.
- [17] Y. Hu, Y. Chen, L. Ding et al., "Pathogenic role of diabetesinduced PPAR-α down-regulation in microvascular dysfunction," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 38, pp. 15401–15406, 2013.
- [18] M. Kanehisa, M. Furumichi, M. Tanabe, Y. Sato, and K. Morishima, "KEGG: new perspectives on genomes, pathways, diseases and drugs," *Nucleic Acids Research*, vol. 45, no. D1, pp. D353–D361, 2017.
- [19] D. R. Zerbino, P. Achuthan, W. Akanni et al., "Ensembl 2018," *Nucleic Acids Research*, vol. 46, no. D1, pp. D754–D761, 2018.
- [20] T. Katome, K. Namekata, Y. Mitamura et al., "Expression of intraocular peroxisome proliferator-activated receptor gamma in patients with proliferative diabetic retinopathy," *Journal of Diabetes and its Complications*, vol. 29, no. 2, pp. 275–281, 2015.
- [21] N. Kaur and V. Vanita, "Association analysis of PPARy (p.Pro12Ala) polymorphism with type 2 diabetic retinopathy in patients from North India," *Ophthalmic Genetics*, vol. 38, no. 3, pp. 217–221, 2016.
- [22] Y. Zhang, N. Meng, Z. Lv, H. Li, and Y. Qu, "The gene polymorphisms of UCP1 but not PPAR γ and TCF7L2 are associated with diabetic retinopathy in Chinese type 2 diabetes mellitus cases," *Acta Ophthalmologica*, vol. 93, no. 3, pp. e223–e229, 2015.
- [23] J. Ma, Y. Li, F. Zhou, X. Xu, G. Guo, and Y. Qu, "Meta-analysis of association between the Pro12Ala polymorphism of the peroxisome proliferator–activated receptor-γ2 gene and diabetic retinopathy in Caucasians and Asians," *Molecular Vision*, vol. 18, p. 2352, 2012.
- [24] Y. Wang, X. H. Wang, and R. X. Li, "Interaction between peroxisome proliferator-activated receptor gamma polymorphism and overweight on diabetic retinopathy in a Chinese case-control study," *International Journal of Clinical and Experimental Medicine*, vol. 8, no. 11, pp. 21647–21652, 2015.
- [25] K. Tariq, S. B. Malik, S. H. B. Ali et al., "Association of Pro12Ala polymorphism in peroxisome proliferator activated receptor gamma with proliferative diabetic retinopathy," *Molecular Vision*, vol. 19, pp. 710–717, 2013.
- [26] S. Qi, C. Wang, Y. Zhang, Y. Cheng, S. Wang, and Y. Zhao, "The association of peroxisome proliferator-activated receptor"

 α with diabetic retinopathy, and additional gene-obesity interaction in Chinese type 2 diabetes mellitus patients," *Obesity Research & Clinical Practice*, vol. 10, pp. S103–S109, 2016.

- [27] K. Muranaka, Y. Yanagi, Y. Tamaki et al., "Effects of peroxisome proliferator-activated receptor γ and its ligand on blood-retinal barrier in a streptozotocin-induced diabetic model," *Investigative Opthalmology & Visual Science*, vol. 47, no. 10, p. 4547, 2006.
- [28] A. Tawfik, T. Sanders, K. Kahook, S. Akeel, A. Elmarakby, and M. Al-Shabrawey, "Suppression of retinal peroxisome proliferator-activated receptor γ in experimental diabetes and oxygen-induced retinopathy: role of NADPH oxidase," *Investigative Opthalmology & Visual Science*, vol. 50, no. 2, p. 878, 2009.
- [29] Q. Chen, F. Qiu, K. Zhou et al., "Pathogenic role of microRNA-21 in diabetic retinopathy through downregulation of PPARα," *Diabetes*, vol. 66, no. 6, pp. 1671–1682, 2017.
- [30] L. Ding, R. Cheng, Y. Hu et al., "Peroxisome proliferatoractivated receptor α protects capillary pericytes in the retina," *The American Journal of Pathology*, vol. 184, no. 10, pp. 2709–2720, 2014.
- [31] T. Behl, I. Kaur, H. Goel, and A. Kotwani, "Implications of the endogenous PPAR-gamma ligand, 15-deoxy-delta-12, 14prostaglandin J2, in diabetic retinopathy," *Life Sciences*, vol. 153, pp. 93–99, 2016.
- [32] X. Z. Dou, D. Nath, Y. Shin, J. X. Ma, and A. S. Duerfeldt, "Structure-guided evolution of a 2-phenyl-4-carboxyquinoline chemotype into PPARα selective agonists: new leads for oculovascular conditions," *Bioorganic & Medicinal Chemistry Letters*, vol. 28, no. 16, pp. 2717–2722, 2018.
- [33] S. R. Savage, C. A. Bretz, and J. S. Penn, "RNA-Seq reveals a role for NFAT-signaling in human retinal microvascular endothelial cells treated with TNFα," *PLoS One*, vol. 10, no. 1, article e0116941, 2015.
- [34] Y. Chen, Y. Hu, M. Lin et al., "Therapeutic effects of PPARα agonists on diabetic retinopathy in type 1 diabetes models," *Diabetes*, vol. 62, no. 1, pp. 261–272, 2013.
- [35] M. G. Petrovic, T. Kunej, B. Peterlin, P. Dovc, and D. Petrovic, "Gly482Ser polymorphism of the peroxisome proliferatoractivated receptor-gamma coactivator-1 gene might be a risk factor for diabetic retinopathy in Slovene population (Caucasians) with type 2 diabetes and the Pro12Ala polymorphism of the PPARgamma gene is not," *Diabetes/Metabolism Research and Reviews*, vol. 21, no. 5, pp. 470–474, 2005.
- [36] J. Sajovic, I. Cilenšek, S. Mankoč et al., "Vascular endothelial growth factor (VEGF)-related polymorphisms rs10738760 and rs6921438 are not risk factors for proliferative diabetic retinopathy (PDR) in patients with type 2 diabetes mellitus (T2DM)," *Bosnian Journal of Basic Medical Sciences*, vol. 19, no. 1, pp. 94–100, 2019.