

Review Article

A Role for PPAR β/δ in Ocular Angiogenesis

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The uses of highly selective PPAR β/δ ligands and PPAR β/δ knockout mice have shown a direct ability of PPAR β/δ to regulate angiogenesis in vitro and in vivo in animal models. PPAR β/δ ligands induce the proangiogenic growth factor VEGF in many cells and tissues, though its actions in the eye are not known. However, virtually, all tissue components of the eye express PPAR β/δ . Both angiogenesis and in particular VEGF are not only critical for the development of the retina, but they are also a central component in many common pathologies of the eye, including diabetic retinopathy and age-related macular degeneration, the most common causes of blindness in the Western world. This review, therefore, will discuss the recent evidence of PPAR β/δ -mediated angiogenesis and VEGF release in the context of ocular disorders.

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1. INTRODUCTION

Peroxisome proliferator-activated receptors (PPAR's) belong to the steroid receptor superfamily of ligand-activated transcription factors [1]. Three PPAR's, PPAR α , PPAR β/δ , and PPAR γ , have been identified [2]. PPAR α is predominantly expressed in liver, heart, kidney, brown adipose tissue, and stomach mucosa; PPAR γ is found primarily in adipose tissue; PPAR β/δ is the most ubiquitously expressed [3, 4], though its roles in physiological and pathophysiological processes are far from clear, particularly, in human tissue. The recent development of PPAR β/δ knockout and transgenic mice has started to implicate roles for PPAR β/δ in adipose tissue formation, metabolism, wound healing, brain development, placental function, atherosclerosis, colorectal carcinogenesis, and skeletal muscle function [5–7]. In this review, the emerging role of PPAR β/δ in regulating endothelial function and angiogenesis will be discussed with a particular emphasis to its relevance in the eye.

2. PPAR β/δ LIGANDS

A number of synthetic PPAR β/δ compounds have been described including GW0742X, GW2433, GW9578, L-783,483, GW501516, L-796,449, L-165,461, and compound

F [8, 9]. In addition, putative endogenous PPAR β/δ activators include fatty acids [3, 10], triglycerides [11], the cyclooxygenase (COX) product, prostacyclin [10], the COX/prostacyclin synthase derived endocannabinoid metabolites [12], and *all-trans* retinoic acid (ATRA) [13]. ATRA is derived from vitamin A (retinol) which is found at its highest levels in the eye and is essential for its development and function [14]. Retinol is converted to retinaldehyde, a component of rhodopsin [14] and a functional PPAR γ antagonist [15, 16], which in turn is metabolised to ATRA by retinal dehydrogenases [14]. ATRA has its own family of high-affinity nuclear receptors, the retinoic acid receptor (RAR) α , β , and γ , which like the PPAR's act as heterodimers with RXR α , β , and γ , the receptors for the ATRA isomer 9-*cis* retinoic acid [17]. Although ATRA can activate PPAR β/δ , it is not known which, if any, of its actions are mediated by PPAR β/δ . However, since ATRA is present in such large quantities in ocular tissue, it is potentially an important site for its actions.

3. PPAR β/δ AND ENDOTHELIAL CELLS

Endothelial cells play critical roles in vascular biology, being both the protective inner lining of vessels and the local site for delivery of oxygen to all tissues. Angiogenesis is the process

of new blood vessel/capillary formation from existing vessels, and hypoxia is a major signal which drives the process [18]. PPAR α , PPAR β/δ , and PPAR γ are all expressed in endothelial cells [19]. PPAR α and PPAR γ have well-characterised roles in endothelial cells, both being in general anti-inflammatory, antiproliferative [1], and antiangiogenic in a variety of in vitro and in vivo models, including tumorigenesis [20] and laser-induced retinal injury [21]. In contrast, the role of PPAR β/δ in this important cell type has only recent starting to be elucidated. Initial reports using prostacyclin as a ligand suggested that like PPAR α and PPAR γ , PPAR β/δ promoted endothelial cell apoptosis [22]. In contrast, the use of highly selective synthetic ligands has revealed a contradictory role for PPAR β/δ regulating endothelial cell survival, proliferation, and angiogenesis.

3.1. PPAR β/δ and endothelial cell proliferation and survival

Long- [23] and short-term [24] culture of endothelial cells with the selective ligand GW501516 induces endothelial cell proliferation, an effect associated with the induction of the VEGF receptor (Flt-1; VEGF R1) and VEGF production [23, 24]. In addition to inducing proliferation, PPAR β/δ activation protects cells from oxidant-induced apoptosis. Synthetic PPAR β/δ ligands or activation of the COX-prostacyclin pathway, which signals through PPAR β/δ , induce the endothelial expression of 14-3-3 α protein [25]. 14-3-3 proteins are anti-apoptotic and anti-inflammatory molecules [26]. PPAR β/δ -induced 14-3-3 α blocks oxidant- (H₂O₂-) induced apoptosis by sequestering the proapoptotic protein Bad, stopping its translocation to mitochondrial membranes, where it initiates cytochrome c release and the subsequent activation of the proapoptotic caspase cascade [25].

3.2. PPAR β/δ and angiogenesis

In addition to having effects on endothelial cell proliferation, PPAR β/δ activation potently induces angiogenesis of human vascular endothelial cells in tumour extracellular matrix in vitro and in a murine matrigel plug model in vivo [24]. In addition, the putative PPAR β/δ ligand prostacyclin analogues [27] and ATRA [28] also induce angiogenesis, though the latter appears mostly dependent on its RAR α receptor rather than PPAR β/δ [29]. In human endothelial cells, a major trigger for morphogenesis induced by PPAR β/δ stimulation was the stimulated release of VEGF [24]. In addition to VEGF, mRNA for the matrix metalloproteinase (MMP)-9, a protease important for cell migration was also elevated by PPAR β/δ activation [24]; however, whether this was secondary to VEGF release was not tested. VEGF is expressed as four main splice variants (by amino acid size: VEGF₁₂₁, VEGF₁₆₅, VEGF₁₈₉, VEGF₂₀₆) [29]. VEGF (VEGF-A; VEGF₁₆₅) is a well-characterised central mediator of endothelial cell growth and angiogenesis [29, 30]. Two endothelial VEGF tyrosine kinase receptors have been identified: VEGFR-1/Flt-1, and VEGFR-2/KDR/Flk1. VEGF R2 appears to be the most important receptor in VEGF-induced mitogenesis and permeability [29, 30]. In addition, in two

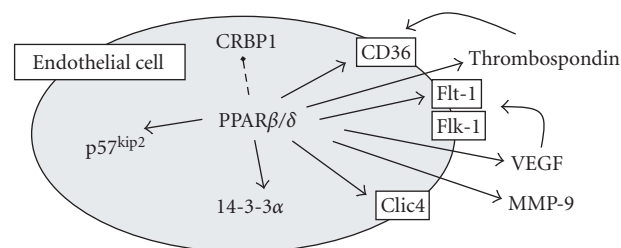


FIGURE 1: Proangiogenic/prosurvival pathways of PPAR β/δ in endothelial cells. PPAR β/δ is expressed in endothelial cells. PPAR β/δ activation induces (solid line) the expression of VEGF and its receptor Flt-1, matrix metalloproteinase (MMP)-9, thrombospondin and its receptor CD36, the chloride intracellular channel protein (CLIC)-4, the cell cycle inhibitor p57^{kip2}, and the antiapoptotic protein 14-3-3 α . In contrast, the cellular retinol binding protein-1 is decreased (dashed line) by PPAR β/δ activation. For those interested, a complex transcriptional map of the potential role of PPAR β/δ as a hub node in tumour angiogenesis has recently also been formed as detailed in [32].

recent studies, the growth of PPAR β/δ wild-type tumours or angiogenesis in matrigel plugs in PPAR β/δ knockout mice was tested [31, 32]. The tumours in PPAR β/δ knockout mice compared to wild-type mice were associated with a diminished blood flow and an immature hyperplastic microvascular structures. Moreover, the retroviral introduction of PPAR β/δ into matrigel plugs was able to rescue the knockout phenotype by triggering microvessel maturation [31]. In the latter of these studies, PPAR β/δ was examined in tumours from patients who had undergone “angiogenic switch” a proangiogenic state involved in tumour progression [32]. PPAR β/δ correlated with advanced pathological tumor stage, increased risk for tumor recurrence, and distant metastasis, and was, therefore, suggested as a hub node transcription factor regulating tumour angiogenesis [32].

Genomic and proteomic analyses of the PPAR β/δ knockout endothelial cells isolated from matrigel plugs have also led to the identification of a number of additional candidate genes to mediate the actions of PPAR β/δ in angiogenesis. In particular, the Cdkn1c gene which encodes the cell cycle inhibitor p57^{kip2} is a direct PPAR β/δ target gene that mediates PPAR β/δ effects on cell morphogenesis [31]. In addition, CD36 and thrombospondin were also decreased in matrigel-invading endothelial cells from PPAR β/δ knockout mice [31]. Thrombospondins by directly interacting with CD36 inhibit angiogenesis in vivo [33, 34]. Similarly, a proteomic analysis by the same group [35] on PPAR β/δ knockout endothelial cells has also revealed a decreased expression of the chloride intracellular channel protein (CLIC)-4 in migrating endothelial cells from PPAR β/δ knockout mice. In contrast, the expression of cellular retinol binding protein CRBP1 is increased in migrating endothelial cells from PPAR β/δ knockout mice [35]. CLIC-4 promotes and plays an essential role during tubular morphogenesis [36], while CRBP1 inhibits cell survival pathways by acting as an inhibitor of the AKT signalling pathway [37], an additional important signalling signal for angiogenesis to occur [38].

The combination of these studies show PPAR β/δ activation induces endothelial cell mitogen and differentiation signals, including VEGF, 14-3-3 α , CD36 and thrombospondin, CLIC4, CRBP-1, and p57^{KIP2}, all of which may act in a coordinate manner to bring about the functional morphogenic changes associated with angiogenesis.

3.3. PPAR β/δ and VEGF

Although the direct evidence for a role of PPAR β/δ in angiogenesis is relatively new, there has been an increasing literature regarding PPAR β/δ regulated tumour cell growth via inducing tumour cells to release VEGF. PPAR β/δ ligands induce VEGF in bladder cancer cells [39], human breast (T47D, MCF7), and prostate (LNCaP, PNT1A) cancer cell lines, along with its receptor flt-1 [22], but not (HT29, colon; HCT116, colon; LS-174T, colon; HepG2, hepatoma; and HuH7, hepatoma) cell lines [40].

In a genetic model of intestinal polyp development APC/min mouse, deletion of PPAR β/δ decreases intestinal adenoma growth and inhibits tumour-promoting effects of the PPAR β/δ agonist GW501516 [41]. Moreover, activation of PPAR β/δ upregulated VEGF in colon carcinoma cells, promoting colon tumour epithelial cell survival through activation of AKT signalling [41]. Angiogenesis was not studied in this model, however, any substantial tumour growth requires a blood supply and angiogenesis to allow it to develop. In contrast, in human colon and liver cancer cell lines [40], PPAR β/δ ligands had no effect on human cancer cell growth, AKT, VEGF or COX-2 expression in vitro or on these markers in the liver, colon, and colon polyps in mice treated in vivo [40]. The roles of PPAR β/δ in VEGF-mediated tumorigenesis are, therefore, still in need of further clarification.

3.4. Expression of PPAR β/δ in the eye

Angiogenesis regulates both the physiological development and many of the most common pathophysiology's of the eye. As yet, there is no direct evidence linking PPAR β/δ and angiogenesis in the eye, however, PPAR β/δ is clearly expressed at least in murine ocular tissue. PPAR β/δ is expressed in the eye ciliary body epithelial cells, cornea epithelial cells, cornea endothelium, cornea fibroblast, retina inner nuclear layer, and retina ganglion cell layer [42]. Although one must be cautious interpreting data from nonocular tissue to the eye [43], as discussed previously and following, pathways that have direct relevance to ocular angiogenesis are clearly regulated by PPAR β/δ and are therefore worthy of discussion.

4. VEGF AND OCULAR ANGIOGENESIS

VEGF is essential in retinal vasculature development [44]. Initially blood vessels grow from the optic nerve outwards. As the retinal tissue develops via a complex interplay between different cellular components such as neurons, glia, endothelial cells, pericytes, and immune cells, the increased oxygen demand induces hypoxia, the main stimulant for new vessel growth via angiogenesis. As the tissue/vasculature develops

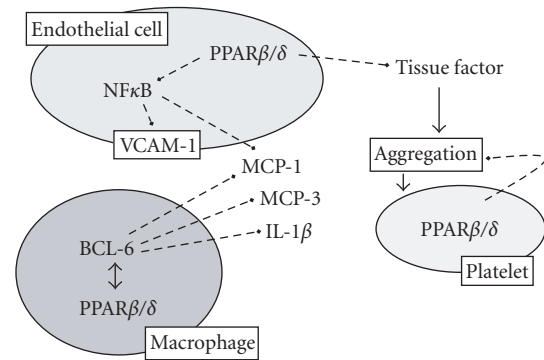


FIGURE 2: Antiinflammatory/anticoagulation pathways of PPAR β/δ . PPAR β/δ activation in endothelial cells reduces NF κ B activation and the induction of vascular cell adhesion molecule (VCAM)-1, and monocyte chemoattractant protein (MCP)-1, along with the release of tissue factor. PPAR β/δ is expressed in platelets and monocytes/macrophages. PPAR β/δ ligands reduce platelet aggregation via a rapid nongenomic mechanism. In macrophages, PPAR β/δ ligands release the transcriptional corepressor BCL-6 from its complex with PPAR β/δ . Free BCL-6 suppresses the release of MCP-1, MCP-3, and IL-1 β .

and gets oxygenated, hypoxia and VEGF decrease limiting new vessel growth [44].

In contrast, neovascularisation of the adult eye via angiogenesis is a critical component of many disorders of the eye including retinopathy of prematurity, diabetic retinopathy, and age-related macular degeneration, the latter two being the leading causes of blindness in the Western world (as reviewed in detail elsewhere [29, 45–48]). Pathological neovascularisation resulting from tissue damage and hypoxia results in a more complex “inflammatory” angiogenesis. These new vessels are often fragile and leaky leading to haemorrhage and vision disturbance and loss. The main trigger for this new vessel growth still appears to be hypoxia induced VEGF expression [29, 45–48]. Angiogenesis is a homeostatic repair mechanism that is required for the reoxygenation of the damaged ischemic tissue [29, 45–48]. The problems that arise with pathologies such as age-related macular degeneration and diabetic retinopathy are that this new vessel growth is leaky and has a critical inflammatory component. VEGF (in particular VEGF A; VEGF₁₆₅) in addition to directly stimulating angiogenesis is also a potent vascular permeability factor and appears to play a role in regulating the local inflammation associated with pathological neovascularisation [49]. VEGF has become a clear therapeutic target for the treatment of angiogenesis in the eye. The clinical importance of VEGF as a target has recently been further demonstrated with the development and use of two new drugs targeting its actions: Macugen (pegaptanib), an aptamer, and Lucentis (ranibizumab), a FAB fragment, from a humanised monoclonal antibody, which both functionally block VEGF. Moreover, Macugen and Lucentis both show clinical efficacy in patients with age-related macular degeneration [50]; especially when treated early and a mature neovasculature has yet to form. These therapies require local delivery by intravitreal

injection, which although having the benefit of overcoming problems such as systemic VEGF blockade, they are clearly still not ideal, and show that new therapies are still required.

5. PPAR β/δ OCULAR ANGIOGENESIS, INFLAMMATION, AND COAGULATION

Angiogenesis associated with pathophysiology is often associated with multiple process such as tissue damage, inflammation, and coagulation. In contrast, developmental angiogenesis may be a simpler hypoxia driven event. Indeed, an inflammatory response is induced by VEGF during pathological but not physiological ischemia-induced retinal angiogenesis [51, 52]. Moreover, specifically blocking inflammatory cytokines monocyte chemoattractant protein-1 and macrophage inflammatory protein-1 α can reduce retinal neovascularisation [53]. Tissue factor is a critical initiator of blood coagulation, and is associated with tumour aggressiveness and angiogenesis in a variety of cancer cells [54], as well as in choroidal neovascularisation where it promotes fibrin formation and the growth of the choroidal angiogenic complex [55]. One important facet of pathological angiogenesis may therefore be this involvement additional pathways, and a complex interplay between processes of tissue damage, hypoxia, inflammation, and coagulation. A long-term therapeutic aim may therefore be to have revascularisation of hypoxic tissue similar to development without these additional inflammatory/coagulation processes.

PPAR β/δ induces VEGF in a number of cell types and induces angiogenesis. Therefore, one may predict that a PPAR β/δ antagonist would be useful to treat or at least test in models of eye disease that involve neovascularisation. However, PPAR β/δ seems consistent with other PPARs in that it also has anti-inflammatory and anticoagulation properties, suggesting that its properties in ocular angiogenesis may be more complex than one would originally predict.

PPAR β/δ activation suppresses endothelial cell tissue factor expression [12]. PPAR β/δ is also expressed in platelets where its ligands reduce platelet aggregation to a variety of stimuli [56]. Similar to PPAR α and PPAR γ , PPAR β/δ ligands are anti-inflammatory in endothelial cells, inhibiting TNF α -induced upregulation of expression of vascular cell adhesion molecule-1, monocyte chemoattractant protein-1, and nuclear factor (NF) κ B translocation [57]. In macrophages, PPAR β/δ controls inflammatory status by its association and disassociation with the transcriptional repressor BCL-6 [58]; in the absence of ligand, PPAR β/δ physically associates with and inhibits this anti-inflammatory BCL-6. When a PPAR β/δ ligand is added, BCL-6 dissociates from PPAR β/δ and represses the inflammation and levels of monocyte chemoattractant protein-1, -3, and IL-1 β [58].

6. CONCLUSION

PPAR β/δ induces angiogenesis and protects endothelial cells from oxidant damage. A common signal for PPAR β/δ activation in endothelial cells or surrounding tissue may be the induction of VEGF. PPAR β/δ is expressed in all tissues in the eye, however its function has yet to be tested in physiologi-

cal processes, development, or pathophysiological disorders. The development of both the eye and common pathological disorders requires angiogenesis, with VEGF being a primary signalling molecule. Blocking PPAR β/δ may therefore provide a new therapy to treat angiogenic eye disorders. The difference between “physiological” and “pathophysiological” angiogenesis may be additional components of inflammation and coagulation. PPAR β/δ ligands reduce inflammation and components of the coagulation cascade. It will be of great interest to test the roles of PPAR β/δ in the eye as a potential proangiogenic stimulus relieving the hypoxia, while potentially still capable of reducing the damaging inflammatory/coagulation signals.

REFERENCES

- [1] L. A. Moraes, L. Piqueras, and D. Bishop-Bailey, “Peroxisome proliferator-activated receptors and inflammation,” *Pharmacology & Therapeutics*, vol. 110, no. 3, pp. 371–385, 2006.
- [2] C. Dreyer, G. Krey, H. Keller, F. Givel, G. Helftenbein, and W. Wahli, “Control of the peroxisomal β -oxidation pathway by a novel family of nuclear hormone receptors,” *Cell*, vol. 68, no. 5, pp. 879–887, 1992.
- [3] S. A. Kliewer, B. M. Forman, B. Blumberg, et al., “Differential expression and activation of a family of murine peroxisome proliferator-activated receptors,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 91, no. 15, pp. 7355–7359, 1994.
- [4] R. Mukherjee, L. Jow, G. E. Croston, and J. R. Paterniti Jr., “Identification, characterization, and tissue distribution of human peroxisome proliferator-activated receptor (PPAR) isoforms PPAR γ 2 versus PPAR γ 1 and activation with retinoid X receptor agonists and antagonists,” *Journal of Biological Chemistry*, vol. 272, no. 12, pp. 8071–8076, 1997.
- [5] H. Vosper, L. Patel, T. L. Graham, et al., “The peroxisome proliferator-activated receptor δ promotes lipid accumulation in human macrophages,” *Journal of Biological Chemistry*, vol. 276, no. 47, pp. 44258–44265, 2001.
- [6] Y. Barak, D. Liao, W. He, et al., “Effects of peroxisome proliferator-activated receptor δ on placentation, adiposity, and colorectal cancer,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 1, pp. 303–308, 2002.
- [7] L. Michalik, B. Desvergne, N. S. Tan, et al., “Impaired skin wound healing in peroxisome proliferator-activated receptor (PPAR) α and PPAR β mutant mice,” *Journal of Cell Biology*, vol. 154, no. 4, pp. 799–814, 2001.
- [8] D. Bishop-Bailey and J. Wray, “Peroxisome proliferator-activated receptors: a critical review on endogenous pathways for ligand generation,” *Prostaglandins & Other Lipid Mediators*, vol. 71, no. 1–2, pp. 1–22, 2003.
- [9] L. Michalik, J. Auwerx, J. P. Berger, et al., “International union of pharmacology. LXI. Peroxisome proliferator-activated receptors,” *Pharmacological Reviews*, vol. 58, no. 4, pp. 726–741, 2006.
- [10] B. M. Forman, J. Chen, and R. M. Evans, “Hypolipidemic drugs, polyunsaturated fatty acids, and eicosanoids are ligands for peroxisome proliferator-activated receptors α and δ ,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 9, pp. 4312–4317, 1997.

- [11] C.-H. Lee, K. Kang, I. R. Mehl, et al., "Peroxisome proliferator-activated receptor δ promotes very low-density lipoprotein-derived fatty acid catabolism in the macrophage," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 7, pp. 2434–2439, 2006.
- [12] M. Ghosh, H. Wang, Y. Ai, et al., "COX-2 suppresses tissue factor expression via endocannabinoid-directed PPAR δ activation," *Journal of Experimental Medicine*, vol. 204, no. 9, pp. 2053–2061, 2007.
- [13] N. Shaw, M. Elholm, and N. Noy, "Retinoic acid is a high affinity selective ligand for the peroxisome proliferator-activated receptor β/δ ," *Journal of Biological Chemistry*, vol. 278, no. 43, pp. 41589–41592, 2003.
- [14] T. Luo, Y. Sakai, E. Wagner, and U. C. Dräger, "Retinoids, eye development, and maturation of visual function," *Journal of Neurobiology*, vol. 66, no. 7, pp. 677–686, 2006.
- [15] O. Ziouzenkova, G. Orasanu, M. Sharlach, et al., "Retinaldehyde represses adipogenesis and diet-induced obesity," *Nature Medicine*, vol. 13, no. 6, pp. 695–702, 2007.
- [16] B. Desvergne, "Retinaldehyde: more than meets the eye," *Nature Medicine*, vol. 13, no. 6, pp. 671–673, 2007.
- [17] M. Mark, N. B. Ghyselinck, and P. Chambon, "Function of retinoid nuclear receptors: lessons from genetic and pharmacological dissections of the retinoic acid signaling pathway during mouse embryogenesis," *Annual Review of Pharmacology and Toxicology*, vol. 46, pp. 451–480, 2006.
- [18] J. Folkman, "Angiogenesis," *Annual Review of Medicine*, vol. 57, pp. 1–18, 2006.
- [19] D. Bishop-Bailey and T. Hla, "Endothelial cell apoptosis induced by the peroxisome proliferator-activated receptor (PPAR) ligand 15-deoxy- $\Delta^{12,14}$ -prostaglandin J_2 ," *Journal of Biological Chemistry*, vol. 274, no. 24, pp. 17042–17048, 1999.
- [20] D. Panigrahy, S. Singer, L. Q. Shen, et al., "PPAR γ ligands inhibit primary tumor growth and metastasis by inhibiting angiogenesis," *Journal of Clinical Investigation*, vol. 110, no. 7, pp. 923–932, 2002.
- [21] T. Murata, S. He, M. Hangai, et al., "Peroxisome proliferator-activated receptor- γ ligands inhibit choroidal neovascularization," *Investigative Ophthalmology & Visual Science*, vol. 41, no. 8, pp. 2309–2317, 2000.
- [22] T. Hatae, M. Wada, C. Yokoyama, M. Shimonishi, and T. Tanabe, "Prostacyclin-dependent apoptosis mediated by PPAR δ ," *Journal of Biological Chemistry*, vol. 276, no. 49, pp. 46260–46267, 2001.
- [23] R. L. Stephen, M. C. U. Gustafsson, M. Jarvis, et al., "Activation of peroxisome proliferator-activated receptor δ stimulates the proliferation of human breast and prostate cancer cell lines," *Cancer Research*, vol. 64, no. 9, pp. 3162–3170, 2004.
- [24] L. Piqueras, A. R. Reynolds, K. M. Hodivala-Dilke, et al., "Activation of PPAR β/δ induces endothelial cell proliferation and angiogenesis," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 27, no. 1, pp. 63–69, 2007.
- [25] J.-Y. Liou, S. Lee, D. Ghelani, N. Matijevic-Aleksic, and K. K. Wu, "Protection of endothelial survival by peroxisome proliferator-activated receptor- δ mediated 14-3-3 up-regulation," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 26, no. 7, pp. 1481–1487, 2006.
- [26] E. Wilker and M. B. Yaffe, "14-3-3 proteins—a focus on cancer and human disease," *Journal of Molecular and Cellular Cardiology*, vol. 37, no. 3, pp. 633–642, 2004.
- [27] R. Pola, E. Gaetani, A. Flex, et al., "Comparative analysis of the in vivo angiogenic properties of stable prostacyclin analogs: a possible role for peroxisome proliferator-activated receptors," *Journal of Molecular and Cellular Cardiology*, vol. 36, no. 3, pp. 363–370, 2004.
- [28] A. Saito, A. Sugawara, A. Uruno, et al., "All-trans retinoic acid induces in vitro angiogenesis via retinoic acid receptor: possible involvement of paracrine effects of endogenous vascular endothelial growth factor signaling," *Endocrinology*, vol. 148, no. 3, pp. 1412–1423, 2007.
- [29] J. Bradley, M. Ju, and G. S. Robinson, "Combination therapy for the treatment of ocular neovascularization," *Angiogenesis*, vol. 10, no. 2, pp. 141–148, 2007.
- [30] A. Hoeben, B. Landuyt, M. S. Highley, H. Wildiers, A. T. Van Oosterom, and E. A. De Bruijn, "Vascular endothelial growth factor and angiogenesis," *Pharmacological Reviews*, vol. 56, no. 4, pp. 549–580, 2004.
- [31] S. Müller-Brüsselbach, M. Kömhoff, M. Rieck, et al., "Deregulation of tumor angiogenesis and blockade of tumor growth in PPAR β -deficient mice," *The EMBO Journal*, vol. 26, no. 15, pp. 3686–3698, 2007.
- [32] A. Abdollahi, C. Schwager, J. Kleeff, et al., "Transcriptional network governing the angiogenic switch in human pancreatic cancer," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 104, no. 31, pp. 12890–12895, 2007.
- [33] M. Febbraio, D. P. Hajjar, and R. L. Silverstein, "CD36: a class B scavenger receptor involved in angiogenesis, atherosclerosis, inflammation, and lipid metabolism," *Journal of Clinical Investigation*, vol. 108, no. 6, pp. 785–791, 2001.
- [34] B. R. Mwaikambo, F. Sennlaub, H. Ong, S. Chemtob, and P. Hardy, "Activation of CD36 inhibits and induces regression of inflammatory corneal neovascularization," *Investigative Ophthalmology & Visual Science*, vol. 47, no. 10, pp. 4356–4364, 2006.
- [35] J. Adamkiewicz, K. Kaddatz, M. Rieck, B. Wilke, S. Müller-Brüsselbach, and R. Müller, "Proteomic profile of mouse fibroblasts with a targeted disruption of the peroxisome proliferator activated receptor- β/δ gene," *Proteomics*, vol. 7, no. 8, pp. 1208–1216, 2007.
- [36] S. Bohman, T. Matsumoto, K. Suh, et al., "Proteomic analysis of vascular endothelial growth factor-induced endothelial cell differentiation reveals a role for chloride intracellular channel 4 (CLIC4) in tubular morphogenesis," *Journal of Biological Chemistry*, vol. 280, no. 51, pp. 42397–42404, 2005.
- [37] Y. S. Kuppumbatti, B. Rexer, S. Nakajo, K. Nakaya, and R. Mira-y-Lopez, "CRBP suppresses breast cancer cell survival and anchorage-independent growth," *Oncogene*, vol. 20, no. 50, pp. 7413–7419, 2001.
- [38] C. D. Kontos, T. P. Stauffer, W.-P. Yang, et al., "Tyrosine 1101 of Tie2 is the major site of association of p85 and is required for activation of phosphatidylinositol 3-kinase and Akt," *Molecular and Cellular Biology*, vol. 18, no. 7, pp. 4131–4140, 1998.
- [39] S. Fauconnet, I. Lascombe, E. Chabannes, et al., "Differential regulation of vascular endothelial growth factor expression by peroxisome proliferator-activated receptors in bladder cancer cells," *Journal of Biological Chemistry*, vol. 277, no. 26, pp. 23534–23543, 2002.
- [40] H. E. Hollingshead, R. L. Killins, M. G. Borland, et al., "Peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) ligands do not potentiate growth of human cancer cell lines," *Carcinogenesis*, vol. 28, no. 12, pp. 2641–2649, 2007.
- [41] D. Wang, H. Wang, Y. Guo, et al., "Crosstalk between peroxisome proliferator-activated receptor δ and VEGF stimulates

- cancer progression,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 50, pp. 19069–19074, 2006.
- [42] H. Higashiyama, A. N. Billin, Y. Okamoto, M. Kinoshita, and S. Asano, “Expression profiling of Peroxisome proliferator-activated receptor- δ (PPAR- δ) in mouse tissues using tissue microarray,” *Histochemistry and Cell Biology*, vol. 127, no. 5, pp. 485–494, 2007.
- [43] P. A. Campochiaro, “Ocular versus extraocular neovascularization: mirror images or vague resemblances,” *Investigative Ophthalmology & Visual Science*, vol. 47, no. 2, pp. 462–474, 2006.
- [44] M. Fruttiger, “Development of the retinal vasculature,” *Angiogenesis*, vol. 10, no. 2, pp. 77–88, 2007.
- [45] J. Chen and L. E. H. Smith, “Retinopathy of prematurity,” *Angiogenesis*, vol. 10, no. 2, pp. 133–140, 2007.
- [46] C. Starita, M. Patel, B. Katz, and A. P. Adamis, “Vascular endothelial growth factor and the potential therapeutic use of pegaptanib (macugen®) in diabetic retinopathy,” *Developments in Ophthalmology*, vol. 39, pp. 122–148, 2007.
- [47] F. Shojaei and N. Ferrara, “Antiangiogenesis to treat cancer and intraocular neovascular disorders,” *Laboratory Investigation*, vol. 87, no. 3, pp. 227–230, 2007.
- [48] U. M. Schmidt-Erfurth, G. Richard, A. Augustin, et al., “Guidance for the treatment of neovascular age-related macular degeneration,” *Acta Ophthalmologica Scandinavica*, vol. 85, no. 5, pp. 486–494, 2007.
- [49] A. P. Adamis and D. T. Shima, “The role of vascular endothelial growth factor in ocular health and disease,” *Retina*, vol. 25, no. 2, pp. 111–118, 2005.
- [50] A. L. Takeda, J. Colquitt, A. J. Clegg, and J. Jones, “Pegaptanib and ranibizumab for neovascular age-related macular degeneration: a systematic review,” *British Journal of Ophthalmology*, vol. 91, no. 9, pp. 1177–1182, 2007.
- [51] S. Ishida, T. Usui, K. Yamashiro, et al., “VEGF₁₆₄-mediated inflammation is required for pathological, but not physiological, ischemia-induced retinal neovascularization,” *Journal of Experimental Medicine*, vol. 198, no. 3, pp. 483–489, 2003.
- [52] T. Usui, S. Ishida, K. Yamashiro, et al., “VEGF₁₆₄₍₁₆₅₎ as the pathological isoform: differential leukocyte and endothelial responses through VEGFR1 and VEGFR2,” *Investigative Ophthalmology & Visual Science*, vol. 45, no. 2, pp. 368–374, 2004.
- [53] S. Yoshida, A. Yoshida, T. Ishibashi, S. G. Elner, and V. M. Elner, “Role of MCP-1 and MIP-1 α in retinal neovascularization during postischemic inflammation in a mouse model of retinal neovascularization,” *Journal of Leukocyte Biology*, vol. 73, no. 1, pp. 137–144, 2003.
- [54] J. Rak, C. Milsom, L. May, P. Klement, and J. Yu, “Tissue factor in cancer and angiogenesis: the molecular link between genetic tumor progression, tumor neovascularization, and cancer coagulopathy,” *Seminars in Thrombosis and Hemostasis*, vol. 32, no. 1, pp. 54–70, 2006.
- [55] H. E. Grossniklaus, J. X. Ling, T. M. Wallace, et al., “Macrophage and retinal pigment epithelium expression of angiogenic cytokines in choroidal neovascularization,” *Molecular Vision*, vol. 8, pp. 119–126, 2002.
- [56] F. Y. Ali, S. J. Davidson, L. A. Moraes, et al., “Role of nuclear receptor signaling in platelets: antithrombotic effects of PPAR β ,” *The FASEB Journal*, vol. 20, no. 2, pp. 326–328, 2006.
- [57] Y. Rival, N. Benéteau, T. Taillandier, et al., “PPAR α and PPAR δ activators inhibit cytokine-induced nuclear translocation of NF- κ B and expression of VCAM-1 in EAhy926 endothelial cells,” *European Journal of Pharmacology*, vol. 435, no. 2-3, pp. 143–151, 2002.
- [58] C.-H. Lee, A. Chawla, N. Urbiztondo, D. Liao, W. A. Boisvert, and R. M. Evans, “Transcriptional repression of atherogenic inflammation: modulation by PPAR δ ,” *Science*, vol. 302, no. 5644, pp. 453–457, 2003.



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