

Corrigendum

Corrigendum to “Development and Characterization of an Electroless Plated Silver/Cysteine Sensor Platform for the Electrochemical Determination of Aflatoxin B₁”

Alex Paul Wacoo ^{1,2}, Mathew Ocheng ³, Deborah Wendiro ¹, Peter California Vuzi ², and Joseph F. Hawumba ²

¹Microbiology and Biotechnology Centre, Department of Product Development, Uganda Industrial Research Institute, P.O. Box 7086, Kampala, Uganda

²Department of Biochemistry and Sports Science, School of Biological Sciences, College of Natural Sciences, Makerere University, P.O. Box 7082, Kampala, Uganda

³Instrumentation Unit, Technology Development Centre, Uganda Industrial Research Institute, P.O. Box 7086, Kampala, Uganda

Correspondence should be addressed to Joseph F. Hawumba; jhawumba@cns.mak.ac.ug

Received 25 November 2020; Accepted 25 November 2020; Published 21 December 2020

Copyright © 2020 Alex Paul Wacoo et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

In the article titled “Development and Characterization of an Electroless Plated Silver/Cysteine Sensor Platform for the Electrochemical Determination of Aflatoxin B₁” [1], anti-Aflatoxin B₁-Peroxidase antibody produced in rabbit IgG fraction of antiserum (product number SAB4200829) (Sigma Aldrich, Saint Louis, MO, USA) was mistakenly used as a reagent instead of anti-aflatoxin B₁ antibody (product number A8679) (Sigma Aldrich, Saint Louis, MO, USA).

Also in the results, Section 3.2 Electrochemical immune detection of aflatoxin B₁ the reading was taken from positive potential which was due to impedance measurement. However, this paper is based on the electro-catalytic activity of horseradish peroxidase on the negative potential. In the method Section 2.3, therefore, entry 600 nm should be -600 nm. In the result section, the whole of Section 3.2 should be revised as follows:

3.2. Performance of the Electrochemical Immunosensor

In order to test the performance of the developed sensor platform for the analysis of aflatoxin B₁, the sensor platform served as a working electrode and a competitive immuno-

sensor format (Figure 1; step 5) was performed both in the absence and presence of free aflatoxin B₁ (Section 2.3). The response of the sensor electrode was tested using different concentrations of aflatoxin B₁ (0-1 ng l⁻¹) and differential staircase voltammetry (DSCV) [20] was used for monitoring the signal (Figure 5). Figure 5(a) shows that the DSCV currents increase with a decrease in aflatoxin B₁ concentrations, suggesting that the peak potential is inversely proportional to aflatoxin B₁ concentration. Since the rate of reaction was dependent on the catalytic activity of HRP, the higher the concentration of bound anti-aflatoxin B₁ antibody-HRP the higher the rate of reaction [2]. Therefore, in the absence of aflatoxin in the solution, more anti-aflatoxin B₁ antibody-HRP bind on the sensor and the current generated was very high. While at high aflatoxin concentration, low level of anti-aflatoxin B₁ antibody-HRP was available to bind on the sensor, and the output current generated was low due to reduced catalytic activity from HRP.

In the Section 3.3, stability and selectivity of the immunosensor entries are 60 mV of the 4160 mV, 360 mV of the 4160 mV, and 760 mV of the 4160 mV should be 10 mV of the 716 mV, 62 mV of the 716 mV, and 130 mV of the 716 mV, respectively. The correct figure is as follows:

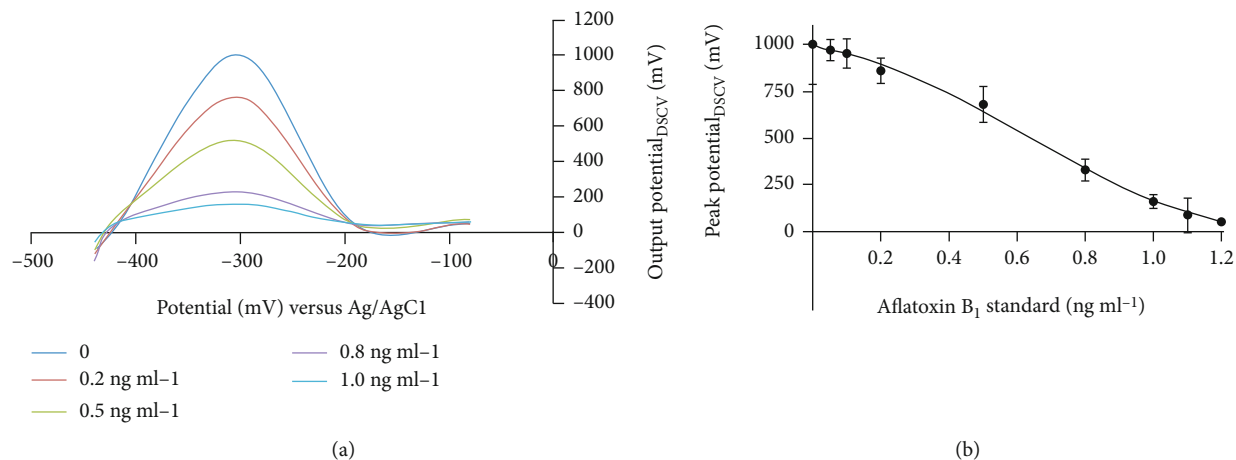


FIGURE 5: (a) DSCV response recorded for silver|cysteine|afatoxin B₁|HRP-blocked immunoelectrode for 0-1 ng ml⁻¹ of aflatoxin B₁ concentrations in a pH 6.5 acetate buffer solution using 0.5 μg ml⁻¹ of anti-aflatoxin B₁ antibody-HRP (scan rate of 20 mV/s). (b) A calibration curve of peak DSCV potential (mV) versus aflatoxin B₁ concentration (ng ml⁻¹).

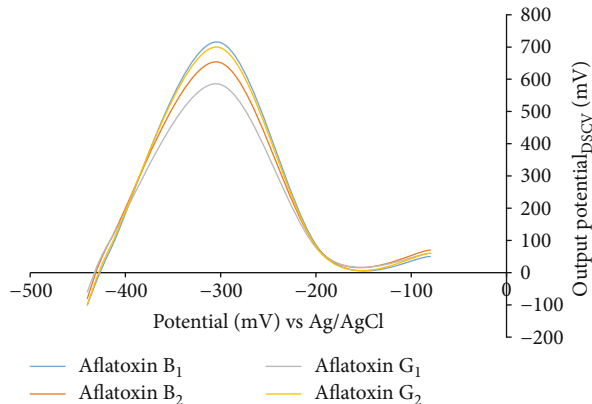


FIGURE 7: The specificity of the immunosensor platform toward aflatoxin 0.2 ng ml⁻¹ B₁, 1.0 ng ml⁻¹ B₂, 1.0 ng ml⁻¹ G₁, and 1.0 ng ml⁻¹ G₂.

Also, due to revision proposed for Section 3.2, Figure 6, Figure 7, and Figure 8 should be numbered as Figure 5, Figure 6, and Figure 7, respectively.

References

- [1] P. A. Wacoo, M. Ocheng, D. Wendi, P. C. Vuzi, and F. J. Hawumba, "Development and characterization of an electrodeless plated silver/cysteine sensor platform for the electrochemical determination of aflatoxin B₁," *Journal of Sensors*, vol. 2016, Article ID 3053019, 8 pages, 2016.
- [2] A. P. Danielson, D. van-Kuren, J. P. Bornstein et al., "Investigating the mechanism of horseradish peroxidase as a raft-initiase," *Polymers*, vol. 10, no. 7, p. 741, 2018.