

Review Article

Early Detection Biomarkers for Ovarian Cancer

**Sreeja Sarojini,¹ Ayala Tamir,¹ Heejin Lim,¹ Shihong Li,² Shifang Zhang,²
Andre Goy,³ Andrew Pecora,³ and K. Stephen Suh^{1,3}**

¹*The Genomics and Biomarker Program, The John Theurer Cancer Center, Hackensack University Medical Center, Hackensack, NJ 07601, USA*

²*Genewiz, Inc., South Plainfield, NJ 07080, USA*

³*The John Theurer Cancer Center, Hackensack University Medical Center, Hackensack, NJ 07601, USA*

Correspondence should be addressed to K. Stephen Suh, ksuh@hackensackumc.org

Received 19 June 2012; Accepted 19 November 2012

Academic Editor: William J. Hoskins

Copyright © 2012 Sreeja Sarojini et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Despite the widespread use of conventional and contemporary methods to detect ovarian cancer development, ovarian cancer remains a common and commonly fatal gynecological malignancy. The identification and validation of early detection biomarkers highly specific to ovarian cancer, which would permit development of minimally invasive screening methods for detecting early onset of the disease, are urgently needed. Current practices for early detection of ovarian cancer include transvaginal ultrasonography, biomarker analysis, or a combination of both. In this paper we review recent research on novel and robust biomarkers for early detection of ovarian cancer and provide specific details on their contributions to tumorigenesis. Promising biomarkers for early detection of ovarian cancer include *KLK6/7*, *GSTT1*, *PRSS8*, *FOLR1*, *ALDH1*, and *miRNAs*.

1. Introduction

Among gynecological malignancies, morbidity and mortality rates are higher among ovarian carcinomas because early detection is difficult due to the absence of recognizable physical symptoms and a lack of sensitive screening methods. In 2012, a total of 22,000 new cases and more than 15,000 deaths are expected, according to Cancer Facts and Figures, 2012, by the American cancer Society [1]. Despite availability of current screening measures, such as transvaginal ultrasound, measurement of biomarker CA125 levels [2], or a combination of both modalities, due to the highly heterogeneous nature of ovarian cancer mortality rates remain high. Although death rate has decreased by 1.9% every year from 2004 to 2008, ovarian cancer still accounts for 3% of all malignancies among women [1]. The long-term survival rate is less than 30% for advanced stage patients, but conventional surgery with chemotherapy can cure about 90% of patients if diagnosed in stage I. Indeed, if the malignancy arises in the ovary and is localized for a sufficient interval to permit effective screening, then the

chances for survival are significantly higher [3]. Because their anatomical location is deep down the pelvis, tumor-related abnormal functioning of the ovaries is asymptomatic until the tumor becomes enlarged or disseminates. In postmenopausal women, the problem is exacerbated because ovaries become dysfunctional after menopause. Therefore, ovarian cancer is more likely to be detected in an advanced rather than an early stage [4]. Microarray analyses and proteomics have been promising technologies used in research to identify molecular signature biomarkers for early detection, disease classification, and prognosis of ovarian cancer. Collections of heterogeneous neoplasms comprising ovarian carcinomas have conventionally been classified based on their type and degree of differentiation. However, current clinical management practices overlook the heterogeneity of ovarian carcinoma [5]. Germline mutations in *BRCA1* and *BRCA2* confer higher risk of ovarian cancer; the estimated risk for *BRCA1* mutation carriers range between 16% and 68% by age 70 and between 11% and 27% for *BRCA2* mutation carriers [6–10]. If diagnosed at a localized stage, the 5 yr survival rate is 93%; however, only 15% of all cases

TABLE 1: Specificity and sensitivity of early detection biomarkers for ovarian cancer from various studies.

Biomarker	Source	<i>n</i> (Total)	Early detection biomarkers: ovarian cancer (sensitivity and specificity)						Reference
			Specificity	Sensitivity	Levels	Benign (<i>n</i>)	Other malignancies (<i>n</i>)	Ovarian (<i>n</i>)	
HE4	Serum	233	95%	73%	High	166	NA	67	[39]
HE4 + CA125	Serum	472	74.20% 76%	100% 92.30%	High	383	89	10%	[36]
Prostasin + CA125	Serum	137	94%	92%	High	100	37	37	[91]
Osteopontin	Plasma	251	NA	NA	High	107	47	51	[138]
KLK6 (hK6)	Serum	384	95%	21%–26%	High	141	NA	146	[55]
KLK6 + CA125	Serum	384	90%	42%	High	141	NA	146	[55]
B7-H4	Serum	2256	97%	45%	High	1023	997	236	[140]
B7-H4 + CA125	Serum	2256	97%	65%	High	1023	410	236	[141]

n: number of patients.

TABLE 2: Clinical trials (currently active or completed) for evaluating novel biomarkers of ovarian cancer.

Biomarker	Condition	Clinical trials for evaluating early detection biomarkers in ovarian cancer (USA)						Primary outcome measure
		Phase	<i>n</i>	Status	Clinical trial no.	Reference		
All biomarkers	Adnexal mass	1	500 (E)	Not yet recruiting	NCT01466049	NA	Screening	
HE4 + CA125	Pelvic mass	0	566	Completed	NCT00315692	[23]	cancer versus benign disease	
CA125	Low risk (w)	1	9500 (E)	Recruiting	NCT00539162	NA	Rate of increase in CA125 over time	
HE4 + CA125	Adnexal mass	1	512	Completed	NCT00987649	NA	Initial cancer risk assessment	
CA125 + HE4	High risk (w)	1	1208 (E)	Recruiting	NCT01121640	NA	PPV of screening protocols	
CA125	High risk (w)	2	2400 (E)	Unknown	NCT00080639	NA	Screening	
Mesothelin	Low risk (w)	0	250 (E)	Unknown	NCT000155740	NA	Screening	
FOLR1	Stage I Ov ca.	2	50 (E)	Recruiting	NCT01511055	NA	Sensitivity and specificity of IOI with folate	
CA125 + TVU	Ovarian diseases	0	750 (E)	Recruiting	NCT01292733	NA	CA125 measurement in blood over time	
CA125 ± TVU	Postmenopausal	0	48230	Completed	NCT00058032	[24, 25]	Screening postmenopausal women	
CA125	High risk (w)	0	2430	Recruiting	NCT00039559	NA	Feasibility at study completion	
CA125 + TVU	High genetic risk (w)	0	5000 (E)	Unknown	NCT00033488	NA	Annual screening	
CA125	High risk (w)	0	6000 (E)	Recruiting	NCT00005095	NA	Screening	
Combined methods	Ov. neoplasms	0	36000	Not yet recruiting	NCT01178736	NA	Low-cost screening	
Interventional	High risk (w)	0	1500	Recruiting	NCT00849199	NA	Genetic testing, screening	
All Biomarkers	High risk (w)	0	250 (E)	Recruiting	NCT00854399	NA	Overall survival	
Tumor markers	High risk (w)	0	5000	Completed	NCT00267072	NA	Early stage detection	
DNA markers	Ovarian cancer	0	170 (E)	Recruiting	NCT00879840	NA	Assessment of screening modalities	
BRCA 1/2 mutation	Ov. neoplasms	0	1500	Completed	NCT00001468	NA	Identifying BRCA 1/2 mutation	

TVU: transvaginal ultrasonography. (w): women, (E): estimated enrollment, IOI: intraoperative imaging. Source: <http://clinicaltrials.gov/>.

are detected at this stage. The majority of cases (63%) are diagnosed after dissemination with the 1-, 5-, and 10-year relative survival rates being 75%, 44%, and 35%, respectively [1]. Clinical trials for identifying *BRCA1* and *BRCA2* mutations in high risk populations are currently being performed (Table 2). As described in earlier reviews, both cytoreductive surgery and combination chemotherapy with platinum-based compounds and taxanes did not change the overall cure rate of ovarian cancer; however, the 5 yr survival rate has increased from 37% (1974–1976) to 46%

(1999–2005) [11]. In order to improve long-term survival of patients, to improve the clinical outcomes of ovarian cancer and to obtain significant reduction of risk, effective early detection methods using screening biomarkers with adequate sensitivity are urgently needed [12].

1.1. CA125 (Cancer Antigen 125). The widely used, classic, “gold standard” tumor biomarker, CA125, a high molecular weight glycoprotein, has a sensitivity between 50% and 60% with a specificity of 90% in early stage postmenopausal

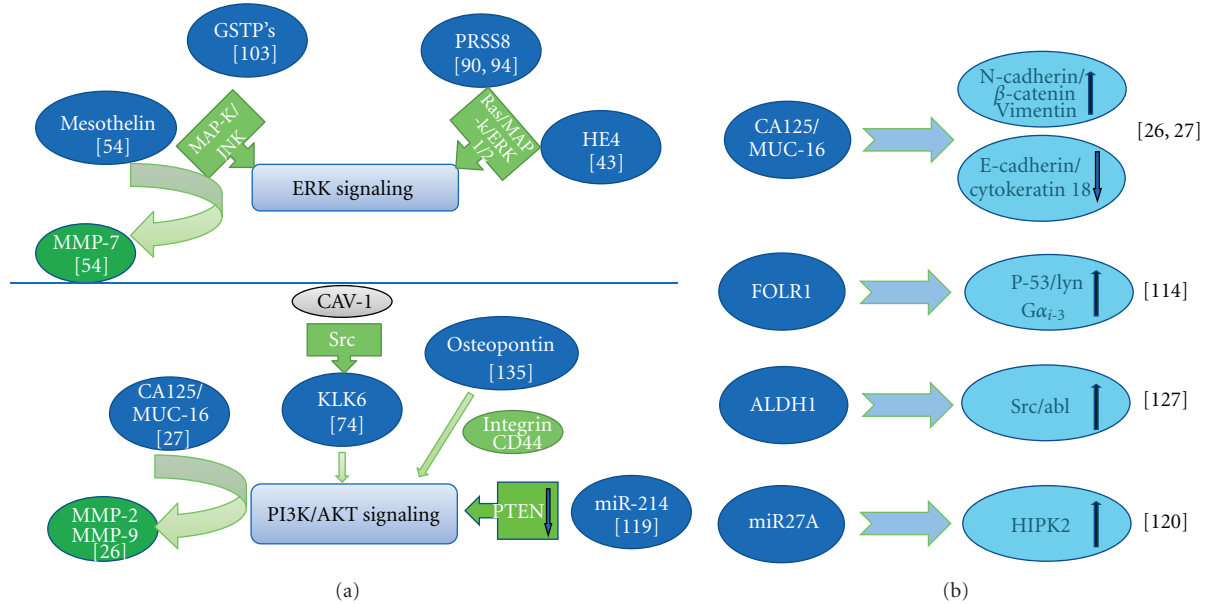


FIGURE 1: EGF/EGFR-based signaling pathways of ovarian cancer biomarkers. (a) Non-EGF/EGFR-based signaling pathways. (b) ↑ indicates upregulation. ↓ indicates downregulation. Caveolin-1 (Cav-1); phosphoinositide-3-kinase (PI3K); c-Jun N-terminal kinase (JNK); extracellular signal-regulated kinase (ERK); homeodomain-interacting protein kinase-2 (HIPK2); matrix metalloproteinase-2 or 7 (MMP-2 or MMP-7); multidrug-resistant protein (MDR1, P-glycoprotein); multidrug resistance-associated proteins 1 and 2 (MRP1/2); mitogen-activated protein kinase (MAP-K); phosphatase and tensin homolog (PTEN).

women, and expression of CA125 is enhanced in 90% of patients with epithelial ovarian cancer above normal levels [13–17]. CA125 is normally expressed in tissues derived from Mullerian and coelomic epithelia and is the only biomarker currently widely used in cancer therapy [18]. It was suggested that CA125 can potentially be used for early detection of ovarian cancer [19] since increased levels of CA125 may precede clinical detection by more than a year. In addition, analysis of CA125 levels has been useful in monitoring chemotherapy responses, distinguishing malignant pelvic masses from benign masses, detection of recurrence, and improving clinical trial design. A decline in expression of CA125 is considered a favorable prognostic occurrence during chemotherapy, and serial measurement of CA125 is used as an indicator of therapeutic outcomes and for assessing stabilization of the disease [15, 20, 21]. However, several factors undermine the significance of CA125 as an early detection biomarker. CA125 expression is absent in about 20% of ovarian cancers, and CA125 expression is elevated in some benign conditions such as liver cirrhosis, endometriosis, and peritonitis. Also, CA125 levels exhibit fluctuations associated with menstrual cycle and pregnancy. As a result, no CA125-based screening techniques are as yet recommended for the general population. However, CA125 has been used effectively in concert with other markers to increase its sensitivity as an early detection biomarker. In a study by Tcherkassova et al., the receptor for circulating fetal protein alpha-fetoprotein (RECAF), an oncofetal antigen, has been examined as a biomarker for early detection of ovarian cancer in conjunction with CA125 among healthy women. When specificity was set at 100% (for each of the

individual markers), it was observed that the addition of RECAF to CA125 enhanced the sensitivity of detection to 83%, as compared to 70% when using CA125 alone. For stages III/IV the sensitivity increased from 79.6% to 88.2% with the addition of RECAF, and a more profound increase was observed for early detection of stages I/II (58.1% with CA125 alone to 76% with RECAF/CA125) [22]. Therefore, because of the relatively low sensitivity of CA125 as a single screening biomarker, combining it with additional biomarkers to create a multiple biomarker panel was more effective; no single biomarker can provide all the necessary information for ovarian cancer diagnosis and therapy. Currently, various clinical trials are evaluating CA125 alone or in combination with other biomarkers for screening of ovarian cancer (Table 2) [23–25]. It was demonstrated that CA125 binds to E-cadherin and β -catenin complexes, which results in enhanced motility, migration, and invasiveness of cells expressing CA125/MUC16 (Figures 1(a) and 1(b)) [26, 27]. As with some other ovarian cancer biomarkers, CA125/MUC16 expressing cells signaling enhance epidermal growth factor receptor (EGFR) activation, which results in increasing its downstream effectors Akt and ERK1/2 and in enhanced MMP-2 and MMP-9 expression [26]. Implementation of computer technology and statistical methods in developing better detection and treatment capacity of ovarian cancer has generated new tools that could boost sensitivity of CA125. One is a computerized algorithm which incorporates and stratifies an individual's age-specific risk for ovarian cancer using CA125 profile; risk of ovarian cancer algorithm (ROCA) increases the sensitivity of CA125 (86%) in preclinical detection. Using ROCA, it could be

predicted whether or not an individual is at high risk based on the levels of CA125 (current and previous) as her age progresses, meaning if the levels of CA125 increase as the individual ages, ROCA identifies the individual as at high risk. Additionally, women with elevated levels of CA125 over 35 u/mL (which is considered as a threshold), which remain unchanged over the years, are identified as at lower risk (specificity 98%). Based on ROCA scores, women are thus triaged into low risk, high risk, and intermediate risk and referred for further procedures such as annuals, transvaginal sonography (TVS), or repeated evaluations of CA125 levels, respectively [28–30]. Similarly, Ova1 is an FDA approved multivariate index for identifying high risk ovarian tumors before any surgical procedures. It combines measurements of five proteins CA125-II, apolipoprotein A1, transthyretin, beta 2 microglobulin, and transferrin. Proprietary OvaCalc software is used to interpret the results, and an Ova1 score will be assigned which varies based on menopausal status. Ova1 score 5 and 4.4 is considered with higher risk of malignancy in premenopausal women and for postmenopausal, respectively, with sensitivity of 92.5% and specificity 42.8% in a trial conducted on women ($n = 516$) referred for surgery by physicians [31]. In a recent study involving 590 women with different types of malignancies including nonepithelial and epithelial ovarian cancers, malignancies metastatic to the ovary, other pelvic cancers, and borderline tumors, Ova1 demonstrated higher sensitivity compared to physician's assessment or to CA125 profile and identified the risk of malignancies when combined with physician assessment before surgery. However, this study demonstrated that Ova1 is independent of cancer stage and menopausal status of women and has high sensitivity in detecting ovarian cancer compared with CA125 and physician assessment [32]. It also demonstrated higher sensitivity in detecting ovarian cancer compared with CA125 alone.

1.2. HE4 (Human Epididymis Protein 4). HE4 is a member of the WFDC family of proteins (whey acidic four-disulfide core) and is found to be overexpressed in ovarian carcinomas. Normal functions of HE4 are yet to be identified; however, the specificity and sensitivity of HE4 shows promise as a serum marker for ovarian cancer in the early detection process [33, 34]. Currently, the FDA has approved the use of HE4 as a tumor marker for monitoring relapse or progression of EOC (epithelial ovarian carcinoma) [35]. Earlier studies evaluated HE4 alone and in combination with CA125 as a biomarker for ovarian cancer. The results suggested that HE4 used in conjunction with CA125 yielded significantly greater specificity than either markers alone [36]. Also, as a single marker, HE4 had the highest sensitivity (72.9% at 95% specificity), and when combined with CA125 sensitivity increased to 76.4% (at 95% specificity). Among biomarkers tested, HE4 levels demonstrated the highest sensitivity for stage I disease, but was only 45.9% at 95% specificity. There was no significant change in sensitivity for stage I disease when HE4 was combined with CA125 or with other biomarkers. Thus, HE4 complements the efficacy of CA125 in improving screening and diagnosis, and together they comprise a promising biomarker panel for detection

and risk stratification of ovarian cancer [35, 37–40]. Recent research by Escudero et al. comparing tumor markers HE4 and CA125 in healthy individuals ($n = 101$), patients with nonmalignant lesions ($n = 535$), and patients with malignant tumors ($n = 423$) indicated that HE4 has higher specificity in patients with benign gynecological disorders than CA125. Similar results were obtained in patients with renal failure or disease. However, the levels of CA125 were higher in all nonovarian malignancies, and the results of this study suggest that even though HE4 has a higher diagnostic specificity than CA125, a combination of both improves the early detection and diagnosis of ovarian cancer of any histological type or stage [41]. In another study conducted among Chinese women ($n = 491$), analysis of HE4 and CA125 in sera from healthy subjects, patients with nonmalignant disorders and ovarian cancer patients showed that both CA125 and HE4 levels were elevated significantly in ovarian cancer patients compared to other groups, with the specificity of HE4 ranging from 90% to 100% and CA125 from 36% (benign gynecologic disease) to 99%, attaining a specificity of 100% for ovarian cancer with the combination of both biomarkers [37]. Furthermore, in a model proposed by Yurkovetsky et al. [42], a multibiomarker panel with CA125, HE4, CEA, and VCAM-1 was highly recommended for early detection of ovarian cancer with 86% sensitivity and 98% specificity. Overall, available data indicates that HE4 could be a novel biomarker for early detection of ovarian cancer in high risk populations, and a multibiomarker panel with CA125 would be promising in detection, diagnosis, and prognosis. HE4 was shown to induce tumor cell adhesion, migration, and growth through the EGFR-MAPK signaling pathway (Figure 1(a)) [43]. In a recent attempt to obtain a better detection tool, serum levels of HE4 and CA125 were incorporated with menopausal status leading to the development of ROMA (risk of ovarian malignancy algorithm) in detecting ovarian cancer from benign pelvic masses even in early stages. ROMA stratifies these patients as high risk groups or low risk, based on ROMA score (numerical) calculated from the predictive index [44]. Recent studies demonstrated that ROMA exhibits high diagnostic accuracy in predicting epithelial ovarian cancer from pelvic masses. However, further research is required for evaluating ROMA in early detection of ovarian cancer [45, 46].

1.3. Mesothelin. Several studies have demonstrated overexpression of mesothelin (a glycoprotein present on mesothelial cells lining the pleura, peritoneum, and pericardium) in most epithelial ovarian cancers and have suggested the eligibility of mesothelin as a target for cancer therapy [47, 48]. Previously Scholler et al. demonstrated that cancer cells undergo CA125/mesothelin dependent cell adhesion in the mesothelial epithelium of peritoneum and confirmed CA125 and mesothelin mediate cell attachment [49]. Rump et al. reported that, this mesothelin/CA125 interaction may also play a role in peritoneal metastasis of ovarian cancer [50]. In a recent study, Lowe et al. evaluated personal factors such as age, BMI, usage of talc, and smoking that influence the levels of expressions of mesothelin, CA125, and HE4 in high-risk, healthy postmenopausal women and demonstrated that

“age” is a significant predictor in expression of mesothelin and HE4 since levels of these biomarkers were found to be increased in older women. Also, there was inverse correlation between mesothelin levels and BMI of the subjects (>50 yr $n = 120$, <50 yr $n = 130$) [51]. Similarly, a significant increase in levels of mesothelin in sera analyzed in normal subjects, subjects with benign disorders, and subjects with malignant ovarian tumors revealed that mesothelin could be a novel biomarker and that higher levels denote poor overall survival in patients following optimal debulking surgery or who have advanced stage ovarian cancer [52]. Moreover, 42% of patients with early stage ovarian cancer had elevated mesothelin in urine compared to only 12% of patients who had elevated mesothelin in serum, suggesting the potential of mesothelin as an early detection biomarker [53]. Also, McIntosh et al. noted that mesothelin and CA125 as a combined marker provided greater sensitivity for early ovarian cancer diagnosis [19]. Cancer cells overexpressing mesothelin demonstrated enhanced migration and metastasis. These activities were mediated through MMP-7, which is regulated through the ERK1/2, Akt, and JNK pathways. The signaling pathway of mesothelin in ovarian cancer is detailed in Figure 1(a), [54].

1.4. Kallikreins. The human kallikrein (KLK) gene family, localized on chromosome 19q13.4, is composed of 15 genes encoding low molecular mass serine proteases (30 KD) of known or predicted trypsin-like or chymotrypsin-like activity, which dysregulate different types of cancer including ovarian, giving either a favorable or unfavorable prognosis [55–57]. KLKs are translated as proenzymes and are cleaved into proenzymes upon release from the secretion pathway. Processing of the proenzymes into active extracellular KLKs is mediated by KLKs or other proteases [58, 59]. Despite the fact that KLKs are involved in the regulation of many physiological processes, including smooth muscle contractions, hormonal regulation, vascular cell growth/repair, and blood pressure, the role of KLKs in pathogenesis or progression of cancer and diabetes remains unclear. The role of KLKs in controlling cellular processes such as neovascularization, apoptosis, and tumor metastasis by cleavage of growth factors, extracellular matrix, or hormones has been previously reported, and robust arteriogenesis induced by overexpression of hK1 has been recently studied [60–62]. KLKs function in numerous physiological and pathological processes, including hormonal regulation [63], either individually or in pathways. Their genetic polymorphisms including sequence and splice variants are often associated with increased risk for various types of cancers including ovarian, thus revealing the potential role of KLKs as prognostic, diagnostic, and predictive biomarkers. KLK4-8, KLK10-11, and KLK13-15 were shown to be upregulated in ovarian tissue and serum from patients and were upregulated in cell lines at the mRNA and/or protein level. Previous studies reported that KLK4 (hK4) proteins are present in normal prostate tissue and are secreted in seminal plasma; however, higher levels of KLK4 expression are associated with the progression of ovarian cancer, mainly late stage serous

epithelial-derived ovarian carcinomas where hK4 represents a potential biomarker for diagnosis and prognosis [64, 65].

KLK4 and KLK5 were reported to be associated with poor outcome in grade 1 and 2 tumors, indicating their association with aggressive forms of cancer. The association of KLK4 with aggressive cancer was identified in an RT-PCR study of KLK4 expression in 147 ovarian cancer tissue samples [66, 67]. Similar patterns of expression were observed in the levels of KLK5 with higher expression in aggressive serous carcinomas compared to expression in normal ovarian tissues or low grade tumors [68].

As demonstrated in Shan et al., KLK6 was reported to be a novel biomarker for ovarian cancer diagnosis based on the fact that it is associated with late stage, chemotherapy responsive, disease-free survival and serous histotype [69, 70]. KLK6 has been identified as having high potential as a novel biomarker with better specificity than CA125 for early detection of ovarian cancer because it is not elevated in noncancerous tumors [55]. Nonetheless, the diagnostic sensitivity is low compared to the diagnostic sensitivity of CA125. However, when KLK6 is used in combination with CA125, the sensitivity of each of the biomarkers is significantly increased (at 90% specificity, sensitivity is 72% for all patients and 42% in early stage patients) [55]. Using an immune-fluorometric assay KLK6 was found in high concentrations in various body fluids including CSF, breast milk, nipple aspirate fluid, and breast cyst fluid of women and in male and female serum [71, 72]. However, the sensitivity and specificity of both KLK6 and CA125 are ineffective in screening a population for early detection of ovarian cancer [73]. The prognosis for patients with preoperative KLK6 levels >4.4 $\mu\text{g/L}$ in serum is much worse than for patients with lower preoperative KLK6 serum levels. The significance of KLK6 as a prognostic factor is higher than CA125. The extensive and almost exclusive sialylation of KLK6 from malignant ovarian cells suggests that sialylated KLK6 could serve as a novel biomarker for early detection [74]. The signaling pathway of KLK6 in ovarian cancer is given in Figure 1(a), where its expression was found to be upregulated through downstream pathways of k-ras. A component of the plasma membrane Caveolae, CAV-1, was shown to be responsible for KLK6 gene expression and related protein secretion [75].

Another important kallikrein family member, KLK7, a chymotryptic serine protease previously reported to have a role in the desquamation of plantar stratum corneum, catalyzes the degradation of desmosomes in the deeper layers of skin during reconstruction [76], thus playing a pivotal role in cell shedding. Similarly, the presence of KLK7 on the surface of cancer cells suggests that, by digestion of extracellular matrix, KLK7 helps in the shedding of tumor cells and, therefore, in invasion and early metastasis. The significance of KLK7 in ovarian cancer early detection is directly related to its upregulated levels in ovarian cancer cells [77]. In a study of 44 ovarian tumors (12 low malignant and 32 carcinomas), Tanimoto et al. [78] showed that levels of KLK7 mRNA were elevated in 66.7% of low malignant potential tumor cells and in 78.1% of malignant cells, suggesting that the overexpression of KLK7 in ovarian tumors contributes to tumor cell growth and metastasis.

KLK8 is normally expressed in ovaries as well as in adult and fetal kidneys, salivary gland, skin, tonsil, and breast. It is also detected in breast milk and amniotic fluid as well as in CVF, CSE, and ovarian cancer ascites [79].

Analysis of kallikreins 4–8, 10, 11, 13, and 14 levels in effusion supernatants obtained from 221 ovarian cancer samples and nonneoplastic diseases demonstrated that, with the exception of KLK4, all kallikreins were expressed at higher than normal levels in ovarian cancer effusions. Among these, KLK6, KLK7, KLK8, and KLK10 showed the highest statistical significance in ovarian cancer effusions over other cancer groups, suggesting that these kallikreins might be useful biomarkers in differential diagnosis of ovarian cancer [80]. In an analysis of kallikreins 6, 10, CA125, and hemostatic markers and 5-year survival outcome from epithelial ovarian carcinoma, it was found that ovarian carcinoma patients who lived past 60 months shared similar elevated preoperative levels of KLK10 and CA125 seen among benign cyst patients. However, the authors indicated a need for a further enlarged study to confirm these findings [81]. Results from an ovarian cancer xenograft model suggest that KLK10 has a tumor suppressive function [82]. Expression of KLK10 is noted in a variety of tissues, including breast, ovary, colon, prostate, and testes [83, 84].

The observed upregulation of KLKs in ovarian cancer is important for diagnosis, prognosis, and treatment. KLK6, KLK10, and KLK11 may provide novel serological diagnostic markers since their expression levels in serum are significantly higher in ovarian cancer patients than in healthy subjects. Similarly, KLK4 and KLK9 share prognostic value in ovarian cancer, with higher expression of KLK5 correlating with poor prognosis [66]. In recent studies, we used a bioinformatics-guided approach coupled with subsequent screening and validation methods for identifying novel biomarkers for ovarian carcinoma. Our results showed that KLK6 and KLK7 are upregulated in ovarian cancer tissues over other cancer types. Upregulation occurs during early stages and in ovarian carcinomas of low malignancy, and these KLKs are secreted into the blood during tumor progression [85]. Hence, KLK6/7 could be further evaluated as early detection biomarkers.

1.5. PRSS8. Human prostatic (PRSS8), a trypsin-like proteinase (40 KDa) localized on chromosome 16p11.2, was first isolated from seminal fluid and was found to be localized or secreted (or both bound and secreted) on the apical surface of the epithelia of the lung, kidney, and prostate. Prostatic plays a significant role in activating epithelial sodium channels and suppressing the in vitro invasiveness of both prostate and breast cancers [86–88]. Similarly, epidermal tight junction formation and terminal differentiation are connected to the matriptase-prostatic proteolytic pathway [89]. Recent studies showed that EGFR (epidermal growth factor receptor) protein expression and EGF-induced phosphorylation of Erk1/2 (extra cellular signal regulated kinases) were found to be downregulated by prostatic expression in PC-3 prostate cancer cells. Given that prostatic functions in EGFR signal modulation, a recent study concluded that it was significant in the regulation of placental trophoblast cell proliferation

via the EGFR-MAPK signaling pathway, since this cascade regulates placental cytotrophoblast proliferation [90].

The potential of prostatic/PRSS8 as a novel biomarker for ovarian carcinoma was suggested by Mok et al. using microarray technology to identify upregulated genes for secretor proteins. The results demonstrated overexpression of PRSS8 in malignant ovarian epithelial cells and stroma compared to the normal ovarian tissue with sensitivity and specificity of 92% and 94%, respectively [91]. A significant decline in postoperative serum levels of PRSS8 was observed in a majority of cases. Similarly, Costa et al. demonstrated significantly higher over-expression of prostatic mRNA in fresh-frozen ovarian cancer tissues than in normal controls [92]. Previous studies to determine the function of Zinc-finger protein 217 (ZNF217) using Affymetrix Gene Chip analysis in the ovarian cancer cell line, HO-8910, with HG-U133 plus 2.0 arrays demonstrated that silencing of the ZNF217 gene resulted in downregulation (approximately 8-fold) of 164 genes compared to normal cells. The same study also confirmed downregulation of PRSS8 after silencing ZNF217 expression indicating the significance of ZNF217 as a key regulator [93] and suggesting PRSS8 as a potential biomarker in ovarian carcinomas. The signaling pathway of PRSS8 in ovarian cancer is detailed in Figure 1(a) [90, 94].

1.6. Glutathione S-Transferase Polymorphisms. Functional polymorphisms of members of the Glutathione S-transferase family (GSTM1, GSTT1, and GSTP1) are the result of large deletions present in the structural gene, which in turn affect drug metabolism and influence the effects of chemotherapy in cancer patients. Allelic variants of GSTs catalyze the conjugation of glutathione to xenobiotic or endogenous substrates, including potentially toxic chemical compounds, and promote detoxification. Given that GST polymorphisms are highly expressed in the human ovary [95] and that polymorphisms of drug metabolizing enzymes influence the susceptibility to different types of cancer, studies on the role of GST polymorphisms in the response to chemotherapy in ovarian cancer therapy would be appropriate. Earlier epidemiologic studies did not confirm the association of GST polymorphisms with epithelial ovarian cancer [96], although they suggested that individuals with homozygous deletions of GSTM or GSTT have reduced or no GST activity, making elimination of electrophilic carcinogens difficult. In a study conducted by Beeghly et al. using DNA extracts from 215 primary epithelial ovarian cancer tissues, GSTT1, GSTM1, and GSTP1 genotypes were identified and assessed by multiplex PCR and PCR-RFLP. The study incorporated Cox proportional hazards regression to determine the association between GST polymorphisms and cancer progression. The results indicated that although none of the individual GST polymorphisms were associated with disease characteristics, when adjusted for disease stage or limited to late-stage patients, GSTM1 polymorphism conferred a better survival. More significantly, combination of no GSTM1 and low GSTP1 resulted in over 60% better progression-free survival and nearly 40% improved overall survival. Therefore, functional polymorphisms of GSTM1 and GSTP1 have important roles in survival of the patients [97]. Similarly,

a meta-analysis, by Economopoulos et al. examining the association of GST polymorphisms and ovarian cancer risk, suggested that GSTT1, GSTM1, and GSTP1 polymorphisms did not seem to contribute any increased risk in individuals. The study included 2357 cases and 3044 controls (8 studies) of GSTM1 null polymorphism, 1923 cases and 2759 controls (6 studies) of GSTT1 null polymorphism, and 3 studies of GSTP1 Ile105Val. Because the populations studied were largely white, the authors indicated that the results could not be extrapolated to other populations, and further race-specific analyses were needed [98]. The role of GSTs is highly relevant in drug-resistant tumors where higher expression of GSTs could alter regulation of the kinase cascade during drug therapy [99]. Similarly, the imbalance between GSH and related enzymes could lead to various pathologies, including cancer, with the genetic polymorphisms of GST affecting susceptibility and progression [99]. Significant reduction in enzymatic activities and higher risk for malignancies are observed in homozygous “null” genotypes (deletion of GSTT1 or GSTM1 genes), because the detoxifying abilities of these individuals are low [100–102]. In ovarian cancer patients with a “double null” genotype, the observed prognosis was poor, along with diminished response to chemotherapy; however, patients with null genotypes for either GSTT1 or GSTM1 exhibited an increased survival rate after chemotherapy for invasive ovarian carcinoma [100–102]. Considering these results, it might be predicted that polymorphisms of GST (GSTT1 or GSTM1) could provide a novel biomarker for early detection and diagnosis of ovarian cancer, although further research is necessary. The signaling pathway of GSTPs in cancer is given in Figure 1(a), [103]. Although it is not yet clear what are the signaling pathways of the different subtypes of GSTP, it was suggested they may operate through the ERK pathway.

1.7. FOLR1. FOLR1 (folate receptor alpha) is a membrane-bound receptor protein involved in transport of folate into cells and other cellular processes. Over-expression of FOLR1 was observed in 69% of uterine serous carcinoma [104]. Rapidly dividing cancer cells have an increased requirement for folate to maintain DNA synthesis, and as reviewed by Kelemen, the expression of FOLR1 is regulated by depletion of extracellular folate levels, accumulation of homocysteine, steroid hormone levels, genetic mutations, and certain transcription factors and cytosolic proteins [105]. Kelemen discusses the significance of folate levels in tumor etiology and progression, with suggestions for future research in FOLR1 gene expression and regulation [105]. Similarly, the over-expression of FOLR1 in various nonmucinous tumors of epithelial origin, including ovarian carcinoma, has been reported; however, its evaluation as a novel biomarker for early detection has yet to be confirmed. FOLR1 over-expression was confirmed in serous ovarian carcinoma in previous studies detailing clinicopathologic features and outcomes, as well as the relationship between FOLR1 and chemoresistance [106]. This study evaluated 91 specimens of serous ovarian carcinomas, and the results showed that over-expression of FOLR1 is a poor prognostic factor for disease-free survival and has a negative impact on overall survival of

patients. Moreover, FOLR1 regulated the expression of bcl-2 and Bax and inhibited cytotoxic drug-induced apoptosis in *in vitro* apoptosis experiments. The results further support that FOLR1 could be a potential biomarker in detection, prognosis, and assessing chemotherapy responses of ovarian carcinoma [106]. In a recent study by van Dam et al., expression of folate receptor-alpha was further examined by using a detecting imaging agent, and intraoperative use of a folate-targeted fluorescence agent with fluorescence microscopy showed a strong signal for all folate-positive malignant tumors and no signal for all folate-negative malignant tumors or benign lesions [107].

Similarly, analysis of the diagnostic and prognostic role of FOLR1 and FOLR3 in effusion cytology of ovarian cancer ($n = 71$), breast cancer ($n = 10$), and malignant mesothelioma ($n = 10$) using quantitative PCR and flow cytometry showed significantly higher concentrations of FOLR1 and FOLR3 in ovarian carcinoma samples compared to breast or mesothelioma. Furthermore, the high expression of folate receptors in ovarian carcinomas shown in this study supports the validity of FOLR1 as drug targets in chemotherapy of ovarian cancer, since FOLR1 expression effectively differentiates ovarian cancer tumors with its coexpression with FOLR3, affecting the serosal cavities of tumors [108]. An earlier study to evaluate the significance of expression of folate receptors in gynecologic tissues (ovary, uterus, and cervix) by Wu et al. revealed contrasting expression patterns of FOLR1 between normal differentiation and malignant transformations of these tissues using quantitative analysis of FOLR1 mRNA. Results indicated that in normal ovary, FOLR1 expression was limited to germinal epithelium, and down-regulation of FOLR1 was noted in differentiation of these cells into benign mucinous or benign serous lesions. Similarly, malignant transformation of these cells also resulted in down regulation of FOLR1, with higher levels of mRNA expression in serous cystadenocarcinoma [109]. Consequently, these studies support the upregulation of FOLR1 in ovarian cancer and confirm that it plays a significant role in regulating folate pathways in the tumor environment, making FOLR1 a possible biomarker for early detection of ovarian carcinoma. Clinical trials are currently being performed to evaluate the potential of FOLR1 as an early detection biomarker (Table 2). The difference in levels of expression of FOLR1 reported in recent studies is summarized in Table 4 [110–113]. Nearly no information is available in regard to the signaling pathway of FOLR1 in ovarian cancer, but it was suggested it signals through p-53/lyn/Gα_{i-3} (Figure 1(b)) [114].

1.8. miRNA. In addition to the above-mentioned biomarkers, epigenetic markers including microRNAs (miRNA) are being considered as positive predictive biomarkers for the clinical management of ovarian cancer [115]. Carcinogenesis is a multistep process involving genetic alterations in oncogenes such as deletions, mutations, or amplifications and changes in microRNA genes. Iorio et al. investigated the importance of miRNA in ovarian cancer and demonstrated that miR-21, miR-141, miR-200a, miR-200c, miR-200b, miR-203, miR-205, and miR-214 could be used as diagnostic markers in ovarian cancer [116]. Taylor et al. compared

TABLE 3: Levels of expression of biomarker ALDH1 in various stages of ovarian cancer.

Biomarker	Expression pattern on tumors	Category	N	Positive rates (levels of expression)	References
ALDH1	Low to high	Serous stages III-IV	65	0% in 27.1% of samples 1%–20% in 44% of samples 20%–100% in 28.9% of samples (10% of all patients demonstrated nearly 100% expression)	[124]
	Low	Malignant tumors	5	17.1% ± 7.61%	[125]
		Benign tumors	5	31.03% ± 6.68%	
		Healthy controls	5	37.4% ± 5.4%	
	Low to high	Serous carcinoma Stage I	266 32	>20% expression in 85% of samples >20% expression in 44% of samples	[121]
	Low and high	Late stage	65	77% positive cells	[126]

TABLE 4: Levels of expression of biomarker FOLR1 in various stages of ovarian cancer.

Biomarker	Expression pattern	Category	N	Positive rates (levels of expression)	References
FOLR1	High	Early stage (I/II)	15	16 ± 2 au	[110]
		Advanced stage (III/IV)	15	12 ± 2 au	
		Healthy controls	30	7 ± 0.9 au	
	High	Advanced stage	104	97%	[111]
		Healthy controls	30	Negligible	
	High	Primary tumors	186	72%	[112]
		Recurrent tumors	27	81.5%	
Weak to moderate	Serous carcinoma	210	81.8%	[113]	
	Nonserous carcinoma	116	39.9%		

these miRNA profiles in circulating tumor exosomes isolated from sera of both benign and malignant ovarian carcinoma patients. The results showed miRNA profiles in exosomal microRNA from ovarian cancer patients were significantly different from the profiles observed in patients with non-malignant disorders, with no exosomal miRNA detected in normal controls [117]. These results suggest that miRNA profiling could be a promising biomarker for early detection of ovarian cancer and biopsy profiling, as well as for screening asymptomatic populations. Further research in OVCAR3 cell lines showed higher levels of miR-21, miR-203, and miR-205 in ovarian cancer compared to normal ovary. miRNA levels were further increased when OVCAR3 cell lines were demethylated with 5-aza-2'-deoxycytidine, suggesting DNA hypomethylation as a possible reason for over-expression of miRNA. This study indicates the pathogenetic role of miRNA in epithelial ovarian cancer and supports miRNA gene methylation as a possible epigenetic pathway for their abnormal expression [116]. In addition, the role of miRNAs in disease prognosis and prediction of outcome in ovarian cancer has also been investigated by profiling miRNA expression from advanced cancer samples [118]. The results indicated that miR-200a, miR-200b, and miR-429 play a role in cancer recurrence and overall survival and demonstrated that low expression of miRNA 200 miRNAs in this group predicts poor outcome, whereas high expression

of miRNA 200 miRNAs inhibits ovarian cancer cell migration, possibly preventing metastasis, which might indicate a better outcome [118]. The results discussed above indicate that miRNAs are aberrantly expressed in ovarian carcinoma and are potential biomarkers for early detection, diagnosis, and monitoring the overall progress of the disease. miR-214 was shown to operate through PI3K/AKT upregulation via PTEN suppression, while it was suggested that miR-27A in ovarian cancer signals through HIPK2 (Figures 1(a) and 1(b)) [119, 120].

1.9. ALDH1 (Aldehyde Dehydrogenase 1). Being a member of aldehyde dehydrogenases protein family, ALDH1A1 plays important role when expressed in a subpopulation of cells with tumor-initiating properties in a variety of malignancies and thus a possible candidate biomarker in cancer therapy. ALDH1 is encoded by ALDH1A1 gene located in chromosome 9q21 and plays key role in pyridine nucleotide-dependent oxidation of aldehydes to respective carboxylic acids. The role of ALDH1 in differentiation of ovarian cancer stem cells and association of ALDH1 expression and various clinicopathologic factors including diagnosis, tumor grade, chemoresponses, staging of disease, and overall survival and disease-free survival of ovarian cancer was evaluated in recent research by Chang et al., using microarray analysis of ALDH1 ($n = 442$) by immune-histochemical staining

as compared to the variations in clinical outcome. Results demonstrated that ALDH1 expression was associated with longer overall survival of the patients, and high expression of ALDH1 is a favorable prognostic factor in patients with ovarian cancer [121]. Similarly, recent study evaluating the expression of ALDH1 in epithelial ovarian cancer stem cells by Steffensen et al. demonstrated the higher expression of ALDH1 in CD44⁺ EOC stem cell clones [122] indicating ALDH1 as a potential biomarker for identifying presence of tumorigenic stem cells and improved therapy options.

Furthermore, tumorigenicity of stem cells coexpressing ALDH1 and CD 133 was studied by Silva et al., who demonstrated that tumor cells coexpressing ALDH1 and CD 133 have highly aggressive phenotype, rapid tumor formation and propagation, worse progression free survival and overall survival in ovarian cancer [123]. Considering these results, which demonstrate that ALDH1A1-positive ovarian cancer cells have increased tumorigenicity and higher chemoresistance, it might be predicted that ALDH1A1, particularly in a marker set, could be a possible biomarker for early detection of ovarian carcinomas [111]. Recently reported levels of expression of ALDH1A1 in various ovarian cancers are detailed in Table 3 [121, 124–126]. The signaling pathway of ALDH1 in ovarian cancer is shown in Figure 1(b) [127].

1.10. Other Relevant Biomarkers. Multianalyte-based analytical discovery platforms readily adaptable to clinical diagnostic screening tests are used currently to profile immune responses against tumor-associated antigens. A goal is to identify tumor-specific antibodies present before the development of clinical symptoms that have potential for detecting ovarian cancer. Such antitumor immune responses are highly beneficial in identifying ovarian cancer [128]. Similarly, tumor vasculature also expresses significant differences from its normal counterpart and is a source of unique markers for detecting various malignancies including ovarian cancer. By using immunohistochemistry-guided laser-capture microdissection and genomewide transcriptional profiling for evaluating the differential expression of genes between tumor cells and normal ovarian tissues, studies have revealed the potential of TVMs (tumor vascular markers) as early detection biomarkers for ovarian cancer [129, 130]. Another potent early detection biomarkers for ovarian cancer are glycans and their associated proteins and lipid structures, which also vary between normal tissue and malignant tumors. Glycosylation is a complex posttranslational modification, and monitoring glycosylation changes provide a more specific and sensitive method for identifying malignancies including ovarian cancer [131, 132]. Microvesicles or exosomes are membranous bodies released from tumor cells and contain macromolecules including RNA, proteins, and lipids. Current research is focusing on identifying tumor exosomes as novel biomarkers for tumor environments since tumor exosomes act as central mediators expressing molecules involved in angiogenesis, stromal remodeling, chemoresistance, activating signaling pathways, and intercellular genetic exchanges [133]. Similarly, the efficiency of FDG-PET/CT (F-18 fluorodeoxyglucose-positron emission tomography) to visualize the increased glucose consumption

of malignant lesions, especially in ovarian cancer, is discussed by Nowosinska et al. In that study, primary malignant tumors could be detected with more accuracy than borderline ovarian tumors; however, limitations included the inability to differentiate between benign and malignant pelvic masses. This may be developed into a potential technique for early detection of ovarian carcinoma and may have application in management of patients [134]. In addition, osteopontin (Figure 1(a)) [135] and B7-H4 have recently been identified as early detection biomarkers for ovarian cancer. These markers are undergoing further research for confirmation [136–138] (Table 1). A recent cell culture study revealed that geometric mean of expression levels of osteopontin in epithelial ovarian cancer cell lines is significantly higher (270.4) than healthy ovarian epithelial cell lines (4.1). Similarly, tissue level expression of osteopontin also varied from normal ovarian epithelial tissue (9) to epithelial ovarian cancer tissue (164). Moreover, immune localization of osteopontin showed higher levels of expression in borderline tumors than benign tumors, suggesting the importance of osteopontin as an early detection biomarker for ovarian cancer [138]. YKL-40, a glycoprotein in chitinase protein family, expresses elevated levels in early and advanced stages of ovarian cancer. Serum levels of YKL-40 from normal healthy individuals, patients at high risk for developing ovarian cancer, and ovarian cancer patients were assessed in a study by Dupont et al. which demonstrated that higher levels of YKL-40 were observed in stage I and stage II patients. Furthermore, YKL-40 levels reliably predicted recurrent and advanced ovarian cancer in these study cohorts since increased levels were observed during advancement of disease [139] indicating that YKL-40 may represent a potential biomarker for early detection of ovarian cancer.

1.11. Genetic Biomarkers. Ovarian cancer, as any other cancers, arises from cells that acquire and accumulate DNA sequence variations. Some of those sequence variations confer the cells a growth advantage and lead to their uncontrolled proliferation (tumorigenesis), unchecked migration (metastasis), and survival against various odds (drug resistance). The advance of sequencing technology is making it possible to uncover those genetic drivers and, thus, identify genetic biomarkers to aid early detection, disease subtyping, staging, and prediction of disease prognosis and selection of effective therapy. The advancement in isolation of small number of circulating tumor cells will eventually make it possible to examine those genetic biomarkers early noninvasively.

As the tip of iceberg, mutations in multiple genes involved in DNA damage repair, cell cycles, cell metabolism, cell adhesions, and other pathways have been reported in association with ovarian cancer. For example, germline mutations in *BRCA1*, *BRCA2*, and *Rad51D* are well known to increase ovarian cancer risk [142, 143]. Whole exome sequencing of 489 high grade serous ovarian cancers (stage II to IV) confirmed the involvement of *BRCA1* and *BRCA2*, with 8%–9% of tumor containing germline mutation and 3% more containing somatic mutation in *BRCA1* and *BRCA2*. The study further identified other recurrently mutated genes,

including *TP53*, *RB1*, *NF1*, *FAT3*, *CSMD3*, *GABRA6*, and *CDK12*. Specifically, 96% of the 489 samples contain mutations in *TP53* [144]. While *TP53* mutations are prevalent in high grade serous cancers, *KRAS* and *BRAF* mutations are more frequent in low-grade subtypes [145]. *CTNNB1* (beta-catenin) mutations are common in endometrioid carcinomas, *PICK3CA* mutations are most frequent in clear cell carcinoma, and *ARID1A* (the AT-rich interactive domain 1A) mutations are often observed in both tumor types [146–148]. We have taken advantage of semiconductor sequencing technology, prepared DNA from 22 serous and endometrioid tumor samples (1 FFPE slide per patient), and sequenced 64 selected genes. With several thousandfold of coverage, we have identified 9 other gene variants that occur in 62%–94% of patients (Li and Suh, *unpublished data*).

Whole transcriptome and exome sequencing revealed that *DICER1* mutations occur at high frequency in non-epithelial ovarian cancers [149]. The mutations are clustered at the metal binding site of the RNase IIIb domain, which are critical for miRNA processing. As reviewed above in section, miRNA themselves are increasingly being considered as biomarkers for ovarian cancer development.

2. Summary

Despite all the conventional and current methods used to detect ovarian cancer development, such as radiographic imaging, invasive biopsies, tumor markers, and a combination of transvaginal ultrasounds with tumor markers, ovarian cancer remains the most common gynecological malignancy and has the highest mortality rate. The identification and validation of early detection biomarkers highly specific to ovarian cancer are needed to establish minimally invasive screening methods for detecting early onset of ovarian cancer. Evaluation of promising biomarkers for early detection opens new horizons in ovarian cancer detection and therapy [150]. The analysis of the human serum proteome has provided better biomarker candidates for early detection, an important goal, as early diagnosis improves the five-year survival rate over 90%. We discuss CA125, a tumor marker with high discriminative power even before the onset of symptoms, which has been demonstrated in many ovarian cancer studies especially in postmenopausal women. We note, however, that the increase in levels of CA125 in other types of cancer, endometriosis, ovulation, other benign ovarian diseases, as well as its low sensitivity in early stages, limits its potential as a single biomarker for ovarian cancer screening. Consequently, a multibiomarker panel aimed at augmenting the sensitivity and specificity of CA125, in which CA125 is used with HE4, mesothelin (Table 1) [141], CEA, VCAM-1, B7-H4, YKL-40, or different combinations is under study for early detection. Of these, HE4 and mesothelin are the most promising candidates to date. Additionally, screening for germline mutations in *BRCA1/BRCA2* is also a promising method for early detection of ovarian cancer in current clinical practice since high risk populations with corresponding mutations could be genetically predisposed toward developing cancer. Prostatein (PRSS8), GSTT1, FOLR1, KLK6, KLK7, and ALDH1 are all currently under

research and clinical trials (Table 2) and are also potential biomarkers for early detection of ovarian cancer. Mok et al. demonstrated the over-expression of PRSS8 in malignant ovarian epithelial cells and stroma with sensitivity and specificity of 92% and 94%, respectively, and with a significant decline in serum postoperative levels. Similarly, evaluation of GST functional polymorphisms (GSTT1, GSTM1) might help in detecting ovarian cancer at early stages, since they affect susceptibility and progression of cancer. However, additional research is needed for confirmation of these possibilities. Also, considering the fact that carriers of low function GST genotypes (GSTT1 null, GSTM1 null) have a strong survival benefit, evaluation of GST polymorphisms could be promising biomarker for early detection of ovarian cancer. Similarly, over-expression of folate receptor-alpha (FOLR1) in 90%–95% nonmucinous tumors of epithelial origin, including epithelial ovarian carcinoma (90%–95%) and serous tumors, indicates the possibility of FOLR1 as an early detection biomarker and suggests the need for further research for confirmation. Several studies demonstrate the significance of KLK6 and KLK7 in ovarian cancer, both being highly expressed in ovarian malignant tumors from early to advanced stages; however, the levels of these proteins in serum samples analyzed at Hackensack University Medical Center had the opposite signature, showing peaks in stage I which declined toward advanced stages [85]. These data support the classification of KLK6/7 as early detection biomarkers. Similarly, small noncoding microRNAs acting as epigenetic regulators cause post transcriptional silencing of target genes and inhibit the activity of antioncogenic pathways promoting tumorigenesis; the aberrant expression of miRNAs has been demonstrated in several studies. Häusler et al. demonstrated higher expression of miR-21, miR-141, miR-200a, miR-200c, miR-200b, miR-203, miR-205, and miR-214 and showed similarity in miRNA profiling in exosomal microRNA from ovarian cancer patients, suggesting that miRNA profiling could be a promising biomarker for early detection of ovarian cancer, biopsy profiling, and for screening asymptomatic populations (Table 1) [140]. Studies have demonstrated that ALDH1-positive ovarian cancer cells have increased tumorigenicity and higher chemoresistance; therefore, it could be predicted that ALDH1, particularly in a marker set, could be a possible biomarker for early detection of ovarian carcinomas (Table 3).

In conclusion, the identification of novel and robust biomarkers with higher specificity and sensitivity for early detection of ovarian cancer could significantly improve the overall survival rate of ovarian cancer patients. The promising biomarkers in this category include KLK6/7, GSTT1, FOLR1, ALDH1, and miRNAs, along with multibiomarker panels in combination with CA125, which is widely used in current practice.

References

- [1] American Cancer Society, *Cancer Facts & Figures 2012*, American Cancer Society, Atlanta, Ga, USA, 2012.

- [2] C. S. Chu and S. C. Rubin, "Screening for ovarian cancer in the general population," *Best Practice and Research Clinical Obstetrics and Gynaecology*, vol. 20, no. 2, pp. 307–320, 2006.
- [3] D. Badgwell and R. C. Bast Jr., "Early detection of ovarian cancer," *Disease Markers*, vol. 23, no. 5-6, pp. 397–410, 2007.
- [4] I. J. Jacobs and U. Menon, "Progress and challenges in screening for early detection of ovarian cancer," *Molecular and Cellular Proteomics*, vol. 3, no. 4, pp. 355–366, 2004.
- [5] K. R. Cho and L. M. Shih, "Ovarian cancer," *Annual Review of Pathology*, vol. 4, pp. 287–313, 2009.
- [6] A. Antoniou, P. D. P. Pharoah, S. Narod et al., "Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case series unselected for family history: a combined analysis of 22 studies," *American Journal of Human Genetics*, vol. 72, no. 5, pp. 1117–1130, 2003.
- [7] A. C. Antoniou, A. P. Cunningham, J. Peto et al., "The BOADICEA model of genetic susceptibility to breast and ovarian cancers: updates and extensions," *British Journal of Cancer*, vol. 98, no. 8, pp. 1457–1466, 2008.
- [8] S. Chen, E. S. Iversen, T. Friebel et al., "Characterization of BRCA1 and BRCA2 mutations in a large United States sample," *Journal of Clinical Oncology*, vol. 24, no. 6, pp. 863–871, 2006.
- [9] S. A. Gayther, W. Warren, S. Mazoyer et al., "Germline mutations of the BRCA1 gene in breast and ovarian cancer families provide evidence for a genotype-phenotype correlation," *Nature Genetics*, vol. 11, no. 4, pp. 428–433, 1995.
- [10] J. P. Struewing, P. Hartge, S. Wacholder et al., "The risk of cancer associated with specific mutations of BRCA1 and BRCA2 among Ashkenazi Jews," *The New England Journal of Medicine*, vol. 336, no. 20, pp. 1401–1408, 1997.
- [11] R. C. Bast Jr., "Biomarkers for ovarian cancer: new technologies and targets to address persistently unmet needs," *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 161–166, 2010.
- [12] K. S. Suh, S. W. Park, A. Castro et al., "Ovarian cancer biomarkers for molecular biosensors and translational medicine," *Expert Review of Molecular Diagnostics*, vol. 10, no. 8, pp. 1069–1083, 2010.
- [13] E. Høgdall, "Cancer antigen 125 and prognosis," *Current Opinion in Obstetrics and Gynecology*, vol. 20, no. 1, pp. 4–8, 2008.
- [14] R. C. Bast Jr., T. L. Klug, E. S. John et al., "A radioimmunoassay using a monoclonal antibody to monitor the course of epithelial ovarian cancer," *The New England Journal of Medicine*, vol. 309, no. 15, pp. 883–887, 1983.
- [15] R. C. Bast Jr., D. Badgwell, Z. Lu et al., "New tumor markers: CA125 and beyond," *International Journal of Gynecological Cancer*, vol. 15, no. 6, supplement 3, pp. 274–281, 2005.
- [16] M. J. Duffy, J. M. Bonfrer, J. Kulpa et al., "CA125 in ovarian cancer: European group on tumor markers guidelines for clinical use," *International Journal of Gynecological Cancer*, vol. 15, no. 5, pp. 679–691, 2005.
- [17] S. Čolaković, V. Lukić, L. Mitrović, S. Jelić, S. Šušnjar, and J. Marinković, "Prognostic value of CA125 kinetics and half-life in advanced ovarian cancer," *International Journal of Biological Markers*, vol. 15, no. 2, pp. 147–152, 2000.
- [18] B. J. D. Rein, S. Gupta, R. Dada, A. Agarwal, J. Safi, and C. Michener, "Potential markers for detection and monitoring of ovarian cancer," *Journal of Oncology*, vol. 2011, Article ID 475983, 17 pages, 2011.
- [19] M. W. McIntosh, C. Drescher, B. Karlan et al., "Combining CA 125 and SMR serum markers for diagnosis and early detection of ovarian carcinoma," *Gynecologic Oncology*, vol. 95, no. 1, pp. 9–15, 2004.
- [20] R. C. Bast Jr., F.-J. Xu, Y.-H. Yu, S. Barnhill, Z. Zhang, and G. B. Mills, "CA 125: the past and the future," *International Journal of Biological Markers*, vol. 13, no. 4, pp. 179–187, 1998.
- [21] A. E. Guppy and G. J. S. Rustin, "CA125 response: can it replace the traditional response criteria in ovarian cancer?" *Oncologist*, vol. 7, no. 5, pp. 437–443, 2002.
- [22] J. Tcherkassova, C. Abramovich, R. Moro et al., "Combination of CA125 and RECAF biomarkers for early detection of ovarian cancer," *Tumor Biology*, vol. 32, no. 4, pp. 831–838, 2011.
- [23] R. G. Moore, D. S. McMeekin, A. K. Brown et al., "A novel multiple marker bioassay utilizing HE4 and CA125 for the prediction of ovarian cancer in patients with a pelvic mass," *Gynecologic Oncology*, vol. 112, no. 1, pp. 40–46, 2009.
- [24] I. Jacobs, A. Gentry-Maharaj, M. Burnell et al., "Sensitivity of transvaginal ultrasound screening for endometrial cancer in postmenopausal women: a case-control study within the UKTOCS cohort," *The Lancet Oncology*, vol. 12, no. 1, pp. 38–48, 2011.
- [25] U. Menon, A. Gentry-Maharaj, R. Hallett et al., "Sensitivity and specificity of multimodal and ultrasound screening for ovarian cancer, and stage distribution of detected cancers: results of the prevalence screen of the UK collaborative trial of ovarian cancer screening (UKTOCS)," *The Lancet Oncology*, vol. 10, no. 4, pp. 327–340, 2009.
- [26] M. Comamala, M. Pinard, C. Thériault et al., "Downregulation of cell surface CA125/MUC16 induces epithelial-to-mesenchymal transition and restores EGFR signalling in NIH:OVCA3 ovarian carcinoma cells," *British Journal of Cancer*, vol. 104, no. 6, pp. 989–999, 2011.
- [27] C. Thériault, M. Pinard, M. Comamala et al., "MUC16 (CA125) regulates epithelial ovarian cancer cell growth, tumorigenesis and metastasis," *Gynecologic Oncology*, vol. 121, no. 3, pp. 434–443, 2011.
- [28] O. Dorigo and J. S. Berek, "Personalizing CA125 levels for ovarian cancer screening," *Cancer Prevention Research*, vol. 4, no. 9, pp. 1356–1359, 2011.
- [29] S. J. Skates, P. Mai, N. K. Horick et al., "Large prospective study of ovarian cancer screening in high-risk women: CA125 cut-point defined by menopausal status," *Cancer Prevention Research*, vol. 4, no. 9, pp. 1401–1408, 2011.
- [30] I. J. Jacobs, S. Skates, A. P. Davies et al., "Risk of diagnosis of ovarian cancer after raised serum CA 125 concentration: a prospective cohort study," *British Medical Journal*, vol. 313, no. 7069, pp. 1355–1358, 1996.
- [31] C. Y. Muller, "Doctor, should I get this new ovarian cancer test-OVA1?" *Obstetrics and Gynecology*, vol. 116, no. 2, pp. 246–247, 2010.
- [32] F. R. Ueland, C. P. Desimone, L. G. Seamon et al., "Effectiveness of a multivariate index assay in the preoperative assessment of ovarian tumors," *Obstetrics and Gynecology*, vol. 117, no. 6, pp. 1289–1297, 2011.
- [33] I. Hellstrom and K. E. Hellstrom, "SMRP and HE4 as biomarkers for ovarian carcinoma when used alone and in combination with CA125 and/or each other," *Advances in Experimental Medicine and Biology*, vol. 622, pp. 15–21, 2008.
- [34] I. Hellström, J. Raycraft, M. Hayden-Ledbetter et al., "The HE4 (WFDC2) protein is a biomarker for ovarian carcinoma," *Cancer Research*, vol. 63, no. 13, pp. 3695–3700, 2003.

- [35] M. Montagnana, E. Danese, S. Giudici et al., "HE4 in ovarian cancer: from discovery to clinical application," *Advances in Clinical Chemistry*, vol. 55, pp. 1–20, 2011.
- [36] R. G. Moore, M. C. Miller, P. Disilvestro et al., "Evaluation of the diagnostic accuracy of the risk of ovarian malignancy algorithm in women with a pelvic mass," *Obstetrics and Gynecology*, vol. 118, no. 2, pp. 280–288, 2011.
- [37] X. Chang, X. Ye, L. Dong et al., "Human epididymis protein 4 (HE4) as a serum tumor biomarker in patients with ovarian carcinoma," *International Journal of Gynecological Cancer*, vol. 21, no. 5, pp. 852–858, 2011.
- [38] J. Li, S. Dowdy, T. Tipton et al., "HE4 as a biomarker for ovarian and endometrial cancer management," *Expert Review of Molecular Diagnostics*, vol. 9, no. 6, pp. 555–566, 2009.
- [39] R. Molina, J. M. Escudero, J. M. Augé et al., "HE4 a novel tumour marker for ovarian cancer: comparison with CA 125 and ROMA algorithm in patients with gynaecological diseases," *Tumour Biology*, vol. 32, no. 6, pp. 1087–1095, 2011.
- [40] R. G. Moore, A. K. Brown, M. C. Miller et al., "The use of multiple novel tumor biomarkers for the detection of ovarian carcinoma in patients with a pelvic mass," *Gynecologic Oncology*, vol. 108, no. 2, pp. 402–408, 2008.
- [41] J. M. Escudero, J. M. Auge, X. Filella, A. Torne, J. Pahisa, and R. Molina, "Comparison of serum human epididymis protein 4 with cancer antigen 125 as a tumor marker in patients with malignant and nonmalignant diseases," *Clinical Chemistry*, vol. 57, no. 11, pp. 1534–1544, 2011.
- [42] Z. Yurkovetsky, S. Skates, A. Lomakin et al., "Development of a multimarker assay for early detection of ovarian cancer," *Journal of Clinical Oncology*, vol. 28, no. 13, pp. 2159–2166, 2010.
- [43] R. Lu, X. Sun, R. Xiao, L. Zhou, X. Gao, and L. Guo, "Human epididymis protein 4 (HE4) plays a key role in ovarian cancer cell adhesion and motility," *Biochemical and Biophysical Research Communications*, vol. 419, no. 2, pp. 274–280, 2012.
- [44] A. J. Li, "New biomarkers for ovarian cancer: OVA1 and ROMA in diagnosis selective use of these new tests may lead to better outcomes for women with adnexal masses or epithelial ovarian cancer," *Contemporary Ob/Gyn*, vol. 57, no. 4, 2012.
- [45] F. Li, R. Tie, K. Chang et al., "Does risk for ovarian malignancy algorithm excel human epididymis protein 4 and ca125 in predicting epithelial ovarian cancer: a meta-analysis," *BMC Cancer*, vol. 12, article 258, 2012.
- [46] M. A. Karlsen, N. Sandhu, C. Høgdall et al., "Evaluation of HE4, CA125, risk of ovarian malignancy algorithm (ROMA) and risk of malignancy index (RMI) as diagnostic tools of epithelial ovarian cancer in patients with a pelvic mass," *Gynecologic Oncology*, vol. 127, no. 2, pp. 379–383, 2012.
- [47] R. Hassan, R. J. Kreitman, I. Pastan, and M. C. Willingham, "Localization of mesothelin in epithelial ovarian cancer," *Applied Immunohistochemistry and Molecular Morphology*, vol. 13, no. 3, pp. 243–247, 2005.
- [48] R. Hassan and M. Ho, "Mesothelin targeted cancer immunotherapy," *European Journal of Cancer*, vol. 44, no. 1, pp. 46–53, 2008.
- [49] N. Scholler, B. Garvik, M. Hayden-Ledbetter, T. Kline, and N. Urban, "Development of a CA125-mesothelin cell adhesion assay as a screening tool for biologics discovery," *Cancer Letters*, vol. 247, no. 1–2, pp. 130–136, 2007.
- [50] A. Rump, Y. Morikawa, M. Tanaka et al., "Binding of ovarian cancer antigen CA125/MUC61 to mesothelin mediates cell adhesion," *The Journal of Biological Chemistry*, vol. 279, no. 10, pp. 9190–9198, 2004.
- [51] K. A. Lowe, C. Shah, E. Wallace et al., "Effects of personal characteristics on serum CA125, mesothelin, and HE4 levels in healthy postmenopausal women at high-risk for ovarian cancer," *Cancer Epidemiology Biomarkers and Prevention*, vol. 17, no. 9, pp. 2480–2487, 2008.
- [52] C. Y. Huang, W. F. Cheng, C. N. Lee et al., "Serum mesothelin in epithelial ovarian carcinoma: a new screening marker and prognostic factor," *Anticancer Research*, vol. 26, no. 6, pp. 4721–4728, 2006.
- [53] D. Badgwell, Z. Lu, L. Cole et al., "Urinary mesothelin provides greater sensitivity for early stage ovarian cancer than serum mesothelin, urinary hCG free beta subunit and urinary hCG beta core fragment," *Gynecologic Oncology*, vol. 106, no. 3, pp. 490–497, 2007.
- [54] M. C. Chang, C. A. Chen, P. J. Chen et al., "Mesothelin enhances invasion of ovarian cancer by inducing MMP-7 through MAPK/ERK and JNK pathways," *Biochemical Journal*, vol. 442, no. 2, pp. 293–302, 2012.
- [55] E. P. Diamandis, A. Scorilas, S. Fracchioli et al., "Human kallikrein 6 (hK6): a new potential serum biomarker for diagnosis and prognosis of ovarian carcinoma," *Journal of Clinical Oncology*, vol. 21, no. 6, pp. 1035–1043, 2003.
- [56] G. M. Yousef and E. P. Diamandis, "Expanded human tissue kallikrein family—a novel panel of cancer biomarkers," *Tumor Biology*, vol. 23, no. 3, pp. 185–192, 2002.
- [57] G. M. Yousef and E. P. Diamandis, "The new human tissue kallikrein gene family: structure, function, and association to disease," *Endocrine Reviews*, vol. 22, no. 2, pp. 184–204, 2001.
- [58] C. A. Borgoño, I. P. Michael, and E. P. Diamandis, "Human tissue kallikreins: physiologic roles and applications in cancer," *Molecular Cancer Research*, vol. 2, no. 5, pp. 257–280, 2004.
- [59] A. Pavlopoulou, G. Pampalakis, I. Michalopoulos, and G. Sotiropoulou, "Evolutionary history of tissue kallikreins," *PLoS ONE*, vol. 5, no. 11, Article ID e13781, 2010.
- [60] O. A. Stone, C. Richer, C. Emanuelli et al., "Critical role of tissue kallikrein in vessel formation and maturation: implications for therapeutic revascularization," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 29, no. 5, pp. 657–664, 2009.
- [61] P. Dominek, P. Campagnolo, M. H. Zadeh et al., "Role of human tissue kallikrein in gastrointestinal stromal tumour invasion," *British Journal of Cancer*, vol. 103, no. 9, pp. 1422–1431, 2010.
- [62] C. A. Borgoño and E. P. Diamandis, "The emerging roles of human tissue kallikreins in cancer," *Nature Reviews Cancer*, vol. 4, no. 11, pp. 876–890, 2004.
- [63] K. Oikonomopoulou, K. K. Hansen, M. Saifeddine et al., "Proteinase-mediated cell signalling: targeting proteinase-activated receptors (PARs) by kallikreins and more," *Biological Chemistry*, vol. 387, no. 6, pp. 677–685, 2006.
- [64] Y. Dong, A. Kaushal, L. Bui et al., "Human kallikrein 4 (KLK4) is highly expressed in serous ovarian carcinomas," *Clinical Cancer Research*, vol. 7, no. 8, pp. 2363–2371, 2001.
- [65] C. V. Obiezu, S. J. C. Shan, A. Soosaipillai et al., "Human kallikrein 4: quantitative study in tissues and evidence for its secretion into biological fluids," *Clinical Chemistry*, vol. 51, no. 8, pp. 1432–1442, 2005.
- [66] C. V. Obiezu, A. Scorilas, D. Katsaros et al., "Higher human kallikrein gene 4 (KLK4) expression indicates poor prognosis of ovarian cancer patients," *Clinical Cancer Research*, vol. 7, no. 8, pp. 2380–2386, 2001.

- [67] C. V. Obiezu and E. P. Diamandis, "Human tissue kallikrein gene family: applications in cancer," *Cancer Letters*, vol. 224, no. 1, pp. 1–22, 2005.
- [68] Y. Dong, A. Kaushal, M. Brattsand, J. Nicklin, and J. A. Clements, "Differential splicing of KLK5 and KLK7 in epithelial ovarian cancer produces novel variants with potential as cancer biomarkers," *Clinical Cancer Research*, vol. 9, no. 5, pp. 1710–1720, 2003.
- [69] S. J. C. Shan, A. Scorilas, D. Katsaros, and E. P. Diamandis, "Transcriptional upregulation of human tissue kallikrein 6 in ovarian cancer: clinical and mechanistic aspects," *British Journal of Cancer*, vol. 96, no. 2, pp. 362–372, 2007.
- [70] N. Emami and E. P. Diamandis, "Utility of kallikrein-related peptidases (KLKs) as cancer biomarkers," *Clinical Chemistry*, vol. 54, no. 10, pp. 1600–1607, 2008.
- [71] E. P. Diamandis, G. M. Yousef, A. R. Soosaipillai et al., "Immunofluorometric assay of human kallikrein 6 (zyme/protease M/neurosin) and preliminary clinical applications," *Clinical Biochemistry*, vol. 33, no. 5, pp. 369–375, 2000.
- [72] E. L. Ashby, P. G. Kehoe, and S. Love, "Kallikrein-related peptidase 6 in Alzheimer's disease and vascular dementia," *Brain Research*, vol. 1363, pp. 1–10, 2010.
- [73] I. J. Jacobs, S. J. Skates, N. MacDonald et al., "Screening for ovarian cancer: a pilot randomised controlled trial," *The Lancet*, vol. 353, no. 9160, pp. 1207–1210, 1999.
- [74] U. Kuzmanov, N. Jiang, C. R. Smith, A. Soosaipillai, and E. P. Diamandis, "Differential N-glycosylation of kallikrein 6 derived from ovarian cancer cells or the central nervous system," *Molecular and Cellular Proteomics*, vol. 8, no. 4, pp. 791–798, 2009.
- [75] R. S. Henkhaus, U. K. B. Roy, D. Cavallo-Medved, B. F. Sloane, E. W. Gerner, and N. A. Ignatenko, "Caveolin-1 mediated expression and secretion of Kallikrein 6 in colon cancer cells," *Neoplasia*, vol. 10, no. 2, pp. 140–148, 2008.
- [76] M. Talieri, E. P. Diamandis, D. Gourgiotis, K. Mathioudaki, and A. Scorilas, "Expression analysis of the human kallikrein 7 (KLK7) in breast tumors: a new potential biomarker for prognosis of breast carcinoma," *Thrombosis and Haemostasis*, vol. 91, no. 1, pp. 180–186, 2004.
- [77] L. G. Kyriakopoulou, G. M. Yousef, A. Scorilas et al., "Prognostic value of quantitatively assessed KLK7 expression in ovarian cancer," *Clinical Biochemistry*, vol. 36, no. 2, pp. 135–143, 2003.
- [78] H. Tanimoto, L. J. Underwood, K. Shigemasa et al., "The stratum corneum chymotryptic enzyme that mediates shedding and desquamation of skin cells is highly overexpressed in ovarian tumor cells," *Cancer*, vol. 86, no. 10, pp. 2074–2082, 1999.
- [79] T. Kishi, L. Grass, A. Soosaipillai, C. Shimizu-Okabe, and E. P. Diamandis, "Human kallikrein 8: immunoassay development and identification in tissue extracts and biological fluids," *Clinical Chemistry*, vol. 49, no. 1, pp. 87–96, 2003.
- [80] L. M. Shih, R. Salani, M. Fiegl et al., "Ovarian cancer specific kallikrein profile in effusions," *Gynecologic Oncology*, vol. 105, no. 2, pp. 501–507, 2007.
- [81] S. C. L. Koh, K. Razvi, Y. H. Chan et al., "The association with age, human tissue kallikreins 6 and 10 and hemostatic markers for survival outcome from epithelial ovarian cancer," *Archives of Gynecology and Obstetrics*, vol. 284, no. 1, pp. 183–190, 2011.
- [82] D. Pépin, Z. Q. Shao, G. Huppé et al., "Kallikreins 5, 6 and 10 differentially alter pathophysiology and overall survival in an ovarian cancer xenograft model," *PLoS ONE*, vol. 6, no. 11, Article ID e26075, 2011.
- [83] E. P. Diamandis, G. M. Yousef, L. Y. Luo, A. Magklara, and C. V. Obiezu, "The new human kallikrein gene family: implications in carcinogenesis," *Trends in Endocrinology and Metabolism*, vol. 11, no. 2, pp. 54–60, 2000.
- [84] H. S. Shvartsman, K. H. Lu, J. Lee et al., "Overexpression of kallikrein 10 in epithelial ovarian carcinomas," *Gynecologic Oncology*, vol. 90, no. 1, pp. 44–50, 2003.
- [85] U. R. Jag, R. Gharbaran, T. Tanaka et al., "Kallikrein family proteases KLK6 and KLK7 are early detection and diagnostic biomarkers for serous and papillary serous subtypes of ovarian cancer," *Cancer Biomarkers*. In press.
- [86] J. X. Yu, L. Chao, and J. Chao, "Prostasin is a novel human serine proteinase from seminal fluid. Purification, tissue distribution, and localization in prostate gland," *The Journal of Biological Chemistry*, vol. 269, no. 29, pp. 18843–18848, 1994.
- [87] J. X. Yu, L. Chao, D. C. Ward, and J. Chao, "Structure and chromosomal localization of the human prostasin (PRSS8) gene," *Genomics*, vol. 32, no. 3, pp. 334–340, 1996.
- [88] G. M. Verghese, M. F. Gutknecht, and G. H. Caughey, "Prostasin regulates epithelial monolayer function: cell-specific Gpld1-mediated secretion and functional role for GPI anchor," *American Journal of Physiology*, vol. 291, no. 6, pp. C1258–C1270, 2006.
- [89] S. Friis, S. Godiksen, J. Bornholdt et al., "Transport via the transcytotic pathway makes prostasin available as a substrate for matriptase," *The Journal of Biological Chemistry*, vol. 286, no. 7, pp. 5793–5802, 2011.
- [90] Y. Y. Fu, W. L. Gao, M. Chen, K. X. Chai, Y. L. Wang, and L. M. Chen, "Prostasin regulates human placental trophoblast cell proliferation via the epidermal growth factor receptor signaling pathway," *Human Reproduction*, vol. 25, no. 3, pp. 623–632, 2010.
- [91] S. C. Mok, J. Chao, S. Skates et al., "Prostasin, a potential serum marker for ovarian cancer: identification through microarray technology," *Journal of the National Cancer Institute*, vol. 93, no. 19, pp. 1458–1464, 2001.
- [92] F. P. Costa, E. L. Batista Jr., A. Zelmanowicz et al., "Prostasin, a potential tumor marker in ovarian cancer—a pilot study," *Clinics*, vol. 64, no. 7, pp. 641–644, 2009.
- [93] G. Sun, J. Qin, Y. Qiu et al., "Microarray analysis of gene expression in the ovarian cancer cell line HO-8910 with silencing of the ZNF217 gene," *Molecular Medicine Reports*, vol. 2, no. 5, pp. 851–855, 2009.
- [94] M. Chen, L. M. Chen, C. Y. Lin, and K. X. Chai, "The epidermal growth factor receptor (EGFR) is proteolytically modified by the Matriptase-Prostasin serine protease cascade in cultured epithelial cells," *Biochimica et Biophysica Acta*, vol. 1783, no. 5, pp. 896–903, 2008.
- [95] M. Rahilly, P. J. Carder, A. Al Nafussi, and D. J. Harrison, "Distribution of glutathione S-transferase isoenzymes in human ovary," *Journal of Reproduction and Fertility*, vol. 93, no. 2, pp. 303–311, 1991.
- [96] S. S. Coughlin and I. J. Hall, "Glutathione S-transferase polymorphisms and risk of ovarian cancer: a HuGE review," *Genetics in Medicine*, vol. 4, no. 4, pp. 250–257, 2002.
- [97] A. Beeghly, D. Katsaros, H. Chen et al., "Glutathione S-transferase polymorphisms and ovarian cancer treatment and survival," *Gynecologic Oncology*, vol. 100, no. 2, pp. 330–337, 2006.
- [98] K. P. Economopoulos, T. N. Sergentanis, and N. F. Vlahos, "Glutathione S-transferase M1, T1, and P1 polymorphisms and ovarian cancer risk: a meta-analysis," *International*

- Journal of Gynecological Cancer*, vol. 20, no. 5, pp. 732–737, 2010.
- [99] D. M. Townsend, K. D. Tew, and H. Tapiero, “The importance of glutathione in human disease,” *Biomedicine and Pharmacotherapy*, vol. 57, no. 3, pp. 145–155, 2003.
- [100] R. E. J. Howells, C. W. E. Redman, K. K. Dhar et al., “Association of glutathione S-transferase GSTM1 and GSTT1 null genotypes with clinical outcome in epithelial ovarian cancer,” *Clinical Cancer Research*, vol. 4, no. 10, pp. 2439–2445, 1998.
- [101] R. E. J. Howells, T. Holland, K. K. Dhar et al., “Glutathione S-transferase GSTM1 and GSTT1 genotypes in ovarian cancer: association with p53 expression and survival,” *International Journal of Gynecological Cancer*, vol. 11, no. 2, pp. 107–112, 2001.
- [102] C. C. McIlwain, D. M. Townsend, and K. D. Tew, “Glutathione S-transferase polymorphisms: cancer incidence and therapy,” *Oncogene*, vol. 25, no. 11, pp. 1639–1648, 2006.
- [103] K. D. Tew, Y. Manevich, C. Grek, Y. Xiong, J. Uys, and D. M. Townsend, “The role of glutathione S-transferase P in signaling pathways and S-glutathionylation in cancer,” *Free Radical Biology and Medicine*, vol. 51, no. 2, pp. 299–313, 2011.
- [104] L. A. Dainty, J. I. Risinger, C. Morrison et al., “Overexpression of folate binding protein and mesothelin are associated with uterine serous carcinoma,” *Gynecologic Oncology*, vol. 105, no. 3, pp. 563–570, 2007.
- [105] L. E. Kelemen, “The role of folate receptor α in cancer development, progression and treatment: cause, consequence or innocent bystander?” *International Journal of Cancer*, vol. 119, no. 2, pp. 243–250, 2006.
- [106] Y. L. Chen, M. C. Chang, C. Y. Huang et al., “Serous ovarian carcinoma patients with high alpha-folate receptor had reducing survival and cytotoxic chemo-response,” *Molecular Oncology*, vol. 6, no. 3, pp. 360–369, 2012.
- [107] G. M. van Dam, G. Themelis, L. M. A. Crane et al., “Intraoperative tumor-specific fluorescence imaging in ovarian cancer by folate receptor- α targeting: first in-human results,” *Nature Medicine*, vol. 17, no. 10, pp. 1315–1319, 2011.
- [108] Y. Yuan, D. A. Nymoen, H. P. Dong et al., “Expression of the folate receptor genes FOLR1 and FOLR3 differentiates ovarian carcinoma from breast carcinoma and malignant mesothelioma in serous effusions,” *Human Pathology*, vol. 40, no. 10, pp. 1453–1460, 2009.
- [109] M. Wu, W. Gunning, and M. Ratnam, “Expression of folate receptor type α in relation to cell type, malignancy, and differentiation in ovary, uterus, and cervix,” *Cancer Epidemiology Biomarkers and Prevention*, vol. 8, no. 9, pp. 775–782, 1999.
- [110] E. Basal, G. Z. Eghbali-Fatourehchi, K. R. Kalli et al., “Functional folate receptor alpha is elevated in the blood of ovarian cancer patients,” *PLoS ONE*, vol. 4, no. 7, Article ID e6292, 2009.
- [111] S. Markert, S. Lassmann, B. Gabriel et al., “Alpha-folate receptor expression in epithelial ovarian carcinoma and non-neoplastic ovarian tissue,” *Anticancer Research*, vol. 28, no. 6A, pp. 3567–3572, 2008.
- [112] K. R. Kalli, A. L. Oberg, G. L. Keeney et al., “Folate receptor alpha as a tumor target in epithelial ovarian cancer,” *Gynecologic Oncology*, vol. 108, pp. 619–626, 2008.
- [113] L. M. Crane, H. J. Arts, M. van Oosten et al., “The effect of chemotherapy on expression of folate receptor-alpha in ovarian cancer,” *Cellular Oncology*, vol. 35, no. 1, pp. 9–18, 2012.
- [114] S. Miotti, M. Bagnoli, A. Tomassetti, M. I. Colnaghi, and S. Canevari, “Interaction of folate receptor with signaling molecules lyn and G α (i-3) in detergent-resistant complexes from the ovary carcinoma cell line IGROV1,” *Journal of Cell Science*, vol. 113, no. 2, pp. 349–357, 2000.
- [115] M. T. M. van Jaarsveld, J. Helleman, E. M. J. J. Berns, and E. A. C. Wiemer, “MicroRNAs in ovarian cancer biology and therapy resistance,” *International Journal of Biochemistry and Cell Biology*, vol. 42, no. 8, pp. 1282–1290, 2010.
- [116] M. V. Iorio, R. Visone, G. Di Leva et al., “MicroRNA signatures in human ovarian cancer,” *Cancer Research*, vol. 67, no. 18, pp. 8699–8707, 2007.
- [117] D. D. Taylor and C. Gercel-Taylor, “MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer,” *Gynecologic Oncology*, vol. 110, no. 1, pp. 13–21, 2008.
- [118] X. Hu, D. M. Macdonald, P. C. Huettner et al., “A miR-200 microRNA cluster as prognostic marker in advanced ovarian cancer,” *Gynecologic Oncology*, vol. 114, no. 3, pp. 457–464, 2009.
- [119] H. Yang, W. Kong, L. He et al., “MicroRNA expression profiling in human ovarian cancer: miR-214 induces cell survival and cisplatin resistance by targeting PTEN,” *Cancer Research*, vol. 68, no. 2, pp. 425–433, 2008.
- [120] Z. Li, S. Hu, J. Wang et al., “MiR-27a modulates MDR1/P-glycoprotein expression by targeting HIPK2 in human ovarian cancer cells,” *Gynecologic Oncology*, vol. 119, no. 1, pp. 125–130, 2010.
- [121] B. Chang, G. Liu, F. Xue et al., “ALDH1 expression correlates with favorable prognosis in ovarian cancers,” *Modern Pathology*, vol. 22, no. 6, pp. 817–823, 2009.
- [122] K. D. Steffensen, A. B. Alvero, Y. Yang et al., “Prevalence of epithelial ovarian cancer stem cells correlates with recurrence in early-stage ovarian cancer,” *Journal of Oncology*, vol. 2011, Article ID 620523, 12 pages, 2011.
- [123] I. A. Silva, S. Bai, K. McLean et al., “Aldehyde dehydrogenase in combination with CD133 defines angiogenic ovarian cancer stem cells that portend poor patient survival,” *Cancer Research*, vol. 71, no. 11, pp. 3991–4001, 2011.
- [124] C. N. Landen Jr., B. Goodman, A. A. Katre et al., “Targeting aldehyde dehydrogenase cancer stem cells in ovarian cancer,” *Molecular Cancer Therapeutics*, vol. 9, no. 12, pp. 3186–3199, 2010.
- [125] K. Pennumatsa, S. L. Edassery, A. Barua, M. J. Bradaric, and J. L. Luborsky, “Differential expression of aldehyde dehydrogenase 1a1 (ALDH1) in normal ovary and serous ovarian tumors,” *Journal of Ovarian Research*, vol. 3, no. 1, article 28, 2010.
- [126] S. Deng, X. Yang, H. Lassus et al., “Distinct expression levels and patterns of stem cell marker, aldehyde dehydrogenase isoform 1 (ALDH1), in human epithelial cancers,” *PLoS ONE*, vol. 5, no. 4, Article ID e10277, 2010.
- [127] J. Kurebayashi, N. Kanomata, T. Moriya, Y. Kozuka, M. Watanabe, and H. Sonoo, “Preferential antitumor effect of the Src inhibitor dasatinib associated with a decreased proportion of aldehyde dehydrogenase 1-positive cells in breast cancer cells of the basal B subtype,” *BMC Cancer*, vol. 10, article 568, 2010.
- [128] M. Chatterjee and M. A. Tainsky, “Autoantibodies as biomarkers for ovarian cancer,” *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 187–201, 2010.
- [129] C. Li, D. Sasaroli, X. Chen et al., “Tumor vascular biomarkers: new opportunities for cancer diagnostics,” *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 253–271, 2010.

- [130] R. J. Buckanovich, D. Sasaroli, A. O'Brien-Jenkins et al., "Tumor vascular proteins as biomarkers in ovarian cancer," *Journal of Clinical Oncology*, vol. 25, no. 7, pp. 852–861, 2007.
- [131] K. L. Abbott, "Glycomic analysis of ovarian cancer: past, present, and future," *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 273–280, 2010.
- [132] H. J. An and C. B. Lebrilla, "A glycomics approach to the discovery of potential cancer biomarkers," *Methods in Molecular Biology*, vol. 600, pp. 199–213, 2010.
- [133] C. D. Roberson, S. Atay, C. Gercel-Taylor, and D. D. Taylor, "Tumor-derived exosomes as mediators of disease and potential diagnostic biomarkers," *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 281–287, 2010.
- [134] E. Nowosinska, S. Avril, I. Murray, T. Szyszko, and N. Avril, "FDG-PET/CT as a molecular biomarker in ovarian cancer," *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 167–175, 2010.
- [135] Y. H. Lin and H. F. Yang-Yen, "The osteopontin-CD44 survival signal involves activation of the phosphatidylinositol 3-kinase/Akt signaling pathway," *The Journal of Biological Chemistry*, vol. 276, no. 49, pp. 46024–46030, 2001.
- [136] I. Simon, D. Katsaros, I. Rigault de la Longrais et al., "B7-H4 is over-expressed in early-stage ovarian cancer and is independent of CA125 expression," *Gynecologic Oncology*, vol. 106, no. 2, pp. 334–341, 2007.
- [137] I. Simon, S. Zhuo, L. Corral et al., "B7-H4 Is a novel membrane-bound protein and a candidate serum and tissue biomarker for ovarian cancer," *Cancer Research*, vol. 66, no. 3, pp. 1570–1575, 2006.
- [138] J. H. Kim, S. J. Skates, T. Uede et al., "Osteopontin as a potential diagnostic biomarker for ovarian cancer," *The Journal of the American Medical Association*, vol. 287, no. 13, pp. 1671–1679, 2002.
- [139] J. Dupont, M. K. Tanwar, H. T. Thaler et al., "Early detection and prognosis of ovarian cancer using serum YKL-40," *Journal of Clinical Oncology*, vol. 22, no. 16, pp. 3330–3339, 2004.
- [140] S. F. M. Häusler, A. Keller, P. A. Chandran et al., "Whole blood-derived miRNA profiles as potential new tools for ovarian cancer screening," *British Journal of Cancer*, vol. 103, no. 5, pp. 693–700, 2010.
- [141] C. A. Shah, K. A. Lowe, P. Paley et al., "Influence of ovarian cancer risk status on the diagnostic performance of the serum biomarkers mesothelin, HE4, and CA125," *Cancer Epidemiology Biomarkers and Prevention*, vol. 18, no. 5, pp. 1365–1372, 2009.
- [142] S. R. Lakhani, S. Manek, F. Penault-Llorca et al., "Pathology of ovarian cancers in BRCA1 and BRCA2 carriers," *Clinical Cancer Research*, vol. 10, no. 7, pp. 2473–2481, 2004.
- [143] C. Loveday, C. Turnbull, E. Ramsay et al., "Germline mutations in RAD51D confer susceptibility to ovarian cancer," *Nature Genetics*, vol. 43, no. 9, pp. 879–882, 2011.
- [144] D. Bell, A. Berchuck, M. Birrer et al., "Integrated genomic analyses of ovarian carcinoma," *Nature*, vol. 474, no. 7353, pp. 609–615, 2011.
- [145] G. Singer, R. Oldt, Y. Cohen et al., "Mutations in BRAF and KRAS characterize the development of low-grade ovarian serous carcinoma," *Journal of the National Cancer Institute*, vol. 95, no. 6, pp. 484–486, 2003.
- [146] J. Palacios and C. Gamallo, "Mutations in the β -catenin gene (CTNNB1) in endometrioid ovarian carcinomas," *Cancer Research*, vol. 58, no. 7, pp. 1344–1347, 1998.
- [147] K. T. Kuo, T. L. Mao, S. Jones et al., "Frequent activating mutations of PIK3CA in ovarian clear cell carcinoma," *American Journal of Pathology*, vol. 174, no. 5, pp. 1597–1601, 2009.
- [148] K. C. Wiegand, S. P. Shah, O. M. Al-Agha et al., "ARID1A mutations in endometriosis-associated ovarian carcinomas," *The New England Journal of Medicine*, vol. 363, no. 16, pp. 1532–1543, 2010.
- [149] A. Heravi-Moussavi, M. S. Anglesio, S.-W. G. Cheng et al., "Recurrent somatic DICER1 mutations in nonepithelial ovarian cancers," *The New England Journal of Medicine*, vol. 366, no. 3, pp. 234–242, 2012.
- [150] B. M. Nolen and A. E. Lokshin, "Screening for ovarian cancer: old tools, new lessons," *Cancer Biomarkers*, vol. 8, no. 4-5, pp. 177–186, 2010.



Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

