

# *Review Article*

# **Balancing Innate Immunity and Inflammatory State via Modulation of Neutrophil Function: A Novel Strategy to Fight Sepsis**

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Sepsis and SIRS (systemic inflammatory response syndrome) belong to a severe disease complex characterized by infection and/or a whole-body inflammatory state. There is a growing body of evidence that neutrophils are actively involved in sepsis and are responsible for both release of cytokines and phagocytosis of pathogens. The neutrophil level is mainly regulated by G-CSF, a cytokine and drug, which is widely used in the septic patient with neutropenia. This review will briefly summarize the role of neutrophils and the therapeutic effect of G-CSF in sepsis. We further suggest that targeting neutrophil function to modulate the balance between innate immunity and inflammatory injury could be a worthwhile therapeutic strategy for sepsis.

## **1. Introduction**

SIRS (systemic inflammatory response syndrome) and sepsis are two different entities of the same disease complex both leading to multiorgan dysfunction and eventually death of the patient.

SIRS is defined as an overwhelming systemic inflammation without infectious component. In contrast, sepsis is a potential fatal medical condition that is characterized by a severe systemic infection accompanied by a dysregulated systemic inflammation [1]. Experimentally, SIRS is often induced by injection of LPS, whereas the frequently used model of sepsis is based on the intra-abdominal inoculation of fecal suspension.

The causes for SIRS and sepsis could be manifold. SIRS can develop as sterile complication of severe trauma, extensive burns, shock, or severe local inflammation such as pancreatitis. The concomitant inflammatory response to the increasing endotoxin levels may result in a vicious cycle leading to SIRS. Sepsis can result from any local or systemic infection and is frequently associated with increased blood levels of endotoxin.

Severe sepsis is a major cause of death in the intensive care unit (ICU) of hospitals and affects millions of people around the world each year. Despite an overwhelming increase in our knowledge regarding the pathogenesis of sepsis and the subsequent advances in clinical care, sepsis still accounts for an unacceptable high mortality ranging from 25 to 30% [2].

## **2. Neutrophils Are the Major Cell Type Involved in SIRS and Sepsis**

*2.1. Neutrophil Is the Primary Line of Defense against Infection.* The neutrophils are the major cell type of the innate immune system, which acts as primary line of defense against invading microbial pathogens [3]. Neutrophils are terminally differentiated hemopoietic cells with a short life span, which respond to infection by migrating from the bloodstream into the infectious site [3].

Efficiency of bacterial elimination is dependent on the rapid recruitment of neutrophils from the circulation, which is promoted by the release of chemotactic agents [4]. Once at the site of infection, neutrophils capture the microbe into a phagosome, which then fuses within transcellular granules forming a phagolysosome. In the phagolysosome, the microbe is destroyed by a combination of oxidative (reactive oxygen species; ROS) and nonoxidative (enzymes, proteases, and antimicrobial peptides) mechanisms [3].

*2.2. The Activation of Neutrophils Induces an Overt Inflammatory Response and Causes Tissue Damage.* Although neutrophils are important for pathogen clearance, the activation of neutrophils also causes an overt inflammatory response and tissue injury. The migration of neutrophils could potentially extend neutrophil-endothelial cell interactions and enhance vascular damage [4]. Local secretion of cytokines by the neutrophils might change the nonthrombogenic properties of endothelial cells to a procoagulant state with the initiation of disseminated intravascular coagulation (DIC) and induce the production of nitric oxide in both endothelial and smooth muscle cells [4]. The inducible nitric oxide (iNOs) is mainly released by neutrophils and has received considerable attention as a mediator of the tissue response to sepsis [5]. The key function of iNOs is to induce the synthesis of nitric oxide (NO), which leads to vasodilation, cytotoxicity, and inflammation [6].

### **3. G-CSF Mobilizes Neutrophils and Enhances the Innate Immunity**

*3.1. Mobilization of Neutrophils Is Induced by Granulocyte Colony-Stimulating Factor (G-CSF).* G-CSF is the principal granulopoietic growth factor regulating the maturation, proliferation, and differentiation of neutrophil precursors and has been used in patients with neutropenia [7]. It enhances maturation of neutrophil functions such as chemotaxis, phagocytes, and bactericidal clearance. It suppresses production of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and promotes the release of IL-1ra and soluble TNF receptor (sTNFr) [8]. It is formed at the site of infection or inflammation but exerts its primary action at a remote organ, that is, bone marrow [8]. G-CSF is able to stimulate the proliferation of neutrophils and plays an important role in modulating the release of inflammatory mediators in acute inflammation.

*3.2. G-CSF Is Modulating the Innate Immune Response.* Several studies have documented the neutrophil-activating effect of G-CSF [9], indicating that it should be considered a potent activator of mature circulating neutrophils. It is capable of priming the respiratory burst, inducing the release of secretory vesicles, and modulating the expression of surface adhesion molecules. The polymorphonuclear surface antigen CD11b/CD18 expression and the plasma elastase- $\alpha$ 1AT complex levels are increased following G-CSF administration.

G-CSF induced mobilization of CD34<sup>+</sup> cells and improved survival of patients with acute-on-chronic liver failure (ACLF) [10]. This effect was attributed to the increased innate immune state, which potentially contributed to prevention from sepsis and multiorgan failure, and improved survival in the study group [11]. A recent experimental study has shown that the use of G-CSF recruited bone marrowderived macrophages into the liver, which, on engraftment in the liver, did help in reducing the hepatic fibrosis and supported hepatic regeneration.

## **4. G-CSF Is Used for Treatment of Septic Patients**

*4.1. G-CSF Treatment Increased Immune Response of the Patients.* According to a large number of clinical trials, G-CSF can decrease the incidence of infections and strengthen host defense in patients. G-CSF administration increased concentrations of IL-1ra, soluble TNF receptors (sTNFr), and IL-10 and reduced TNF- $\alpha$ , IFN- $\gamma$ , and GM-CSF in healthy volunteers [12]. Application of a single dose of G-CSF resulted in the upregulation of neutrophils. These newly formed neutrophils were well equipped against bacterial infections in terms of Fcy RI expression, Fcy RI-dependent antibody-dependent cellular cytotoxicity, and strong-surface CD14 expression. In contrast the biological significance of the decreased surface-expression of Fcy RI11 and the high intracellular LAP content still needs to be elucidated.

*4.2. G-CSF Has Beneficial Effects in Experimental Models of Sepsis.* G-CSF has been used successfully in the past to prevent and to treat experimentally induced sepsis, for example, using the cecal ligation and perforation (CLP) model in rats and mice [13, 14]. It has been reported that the combination of G-CSF application and antibiotic prophylaxis was the most efficacious treatment in rats with polymicrobial peritonitis [15]. Prophylaxis with G-CSF increased the survival rate, decreased the bacterial load, and promoted the production of inflammatory cytokines directly, as demonstrated in experiments [16].

The protective effects were at least partially related to increased neutrophil function, as studies proved that similar phenomena were observed by increasing the neutrophil secretory proteins. Singer observed in a model of iNOs knock-out mice that the iNOs derived NO is a determinant of the proinflammatory phenotype acquired by the hepatic microvasculature during sepsis [5]. Luo et al. found that the neutrophil extracellular trap had a proinflammatory role in abdominal sepsis and regulated the pulmonary infiltration of neutrophils and tissue injury [17].

The results might provide insights and guidance for the application of G-CSF and antibiotic in the perioperative treatment of patients who are susceptible to infection following intra-abdominal surgery.

*4.3. G-CSF Treatment of Patients with Sepsis Leads to Controversial Results.* It has become well accepted that sepsis is composed of two, often concomitant, phases (Figure 1):



FIGURE 1: The development of sepsis has two phases, that is, activating and suppressing phase. The activation of the innate immune system can lead to a balanced response that can trigger the elimination of invading pathogens and the recovery of tissue but can also lead to an unbalanced response that can induce hyperinflammation or immune suppression.

an immune-activated phase and an immune-suppressed phase [1]. Identification of two distinct phases of sepsis calls for a stage dependent therapy. However, it remains difficult since the two stages can be hardly distinguished in experimental models but much less in patient with sepsis.

Therefore, treatment strategies targeting only one phase may fail and cause mortality.

G-CSF is frequently and successfully used in patients with severe neutropenia, who are at risk of sepsis [18]. However, when used in clinical trials to treat ongoing severe sepsis or when given as prophylactic treatment, G-CSF did not result in a clear benefit (Table 1). It is reported that perioperative G-CSF administration was effective in upregulating immune function in patients subjected to major surgery. G-CSF administration resulted in increased levels of natural circulating antagonists of TNF- $\alpha$  and IL-1, that is, TNF-R p55/p75 and IL-1ra in patients, thus increasing the threshold of triggering the inflammatory reaction. Perioperative prophylaxis with G-CSF in high-risk colorectal cancer patients resulted in improved recovery [19]. In acute-on-chronic liver failure (ACLF) patients, treatment of G-CSF significantly decreases the risk of sepsis [10]. Additionally, G-CSF is used in the treatment of neonates and adults with infection [20].

Although G-CSF administration is associated with a longer duration of survival in patients with severe sepsis, a meta-analysis from Bo et al. indicated that G-CSF therapy did not significantly reduce the overall mortality at 14 days or 28 days or in-hospital mortality in patients with sepsis [21]. Nelson et al. reported that G-CSF treatment did effectively increase neutrophil levels but did not affect the mortality rate of the patient with community-acquired pneumonia [22]. A multicenter clinical trial of G-CSF had shown similar results that the G-CSF treatment did not substantially reduce mortality and complication rate in patients with pneumonia and severe sepsis [23]. The clinical trials in nonneutropenic

septic patients indicated that the use of G-CSF decreased the risk of sepsis in nosocomial pneumonia patients, although the effect did not reach statistical significance [24]. Similarly, a clinical observation from melioidosis patients indicated that G-CSF treatment was associated with longer survival duration but was not associated with an overall survival benefit [25]. Another study from the same group showed that the G-CSF treatment did not improve the outcome in patients with septic shock, [26]. Altogether, the G-CSF administration in infectious disease was not associated with a clear therapeutic benefit (Table 1).

## **5. The Therapeutic Effect of G-CSF Might Be Enhanced by Reducing the Inflammatory Response**

*5.1. G-CSF Treatment Induces LPS Sensitization.* Our own results demonstrated that G-CSF pretreatment was not only mobilizing neutrophils but also inducing LPS sensitization to inflammatory response [27]. The inflammatory response contributed to bacterial clearance but did become deleterious if circulating LPS was abundant. Therefore, both components of the disease—the state of innate immunity and the inflammatory response—have to be addressed appropriately to maximize the therapeutic efficiency of GCSF in sepsis.

According to the sepsis treatment guidelines [28–30], therapy consists of elimination of the septic focus by early antibiotic treatment ("hit early, hit hard") as well as supportive therapy consisting of volume resuscitation and maintenance of organ function. Antibiotic treatment is very efficient to reduce the bacterial load. However, the antimicrobial, cytolytic properties of antibiotics may induce the release of LPS from the outer membrane of Gram-negative bacteria [31]. Antibiosis does not address the effect of high circulating



TABLE 1: Clinical studies investigating effect of G-CSF treatment on infectious disease. TABLE 1: Clinical studies investigating effect of G-CSF treatment on infectious disease.

endotoxin levels. Current guidelines do not call for reducing the overwhelming inflammatory response to circulating free endotoxin. New evidence indicated that the controversial clinical observations might be related to the G-CSF modulated inflammatory response.

*5.2. Strategies to Decrease the LPS Induced Inflammatory Response.* A number of strategies exist to reduce circulating LPS levels or to minimize the response to LPS [32]. Elimination of LPS using polymyxin columns is troublesome because of the unproven benefit, significant costs, and potential risks [33]. Neutralization of LPS using antibodies reduced the LPS induced inflammatory response [32]. Currently, vaccination strategies to increase the elimination of pathogens are under investigation [34]. None of these antiendotoxin strategies reached wide clinical acceptance.

Minimizing the response to LPS was explored by blocking different pathways. Blockade of the interaction between LPS and TLR4 signal pathway decreased TNF- $\alpha$  levels in LPS induced SIRS model and *E. coli* induced sepsis model [35]. The intracellular signal transduction cascade can be blocked to prevent the excessive induction of proinflammatory cytokines. However, clinical trials using  $TNF-\alpha$  antibodies were not promising [36].

Recently TLR4 blockade gained increasing attention [37]. The TLR4 signaling pathway leading to LPS-mediated NFkappa B activation constitutes an important therapeutic target for sepsis therapy. Various molecules are involved in regulating TLR4-expression on the cell membrane and act as new adjuvant therapies that are able to weaken the deleterious effects of exaggerated host response to infection [38].

Eritoran tetrasodium is a nonpathogenic endotoxin analog that antagonizes inflammatory signaling via the immune receptor TLR4 [39]. Therefore, eritoran tetrasodium (E5564) was investigated as promising molecular candidate to treat sepsis [40]. Christ et al. proposed that E5531, a slightly different analogue, would antagonize LPS activity at its cellsurface receptor leading to inhibition of transmembrane signal transduction [41]. E5531 protected mice from lethal doses of LPS and from viable *E. coli* infections in combination with antibiotics. However, E5531 did not affect bacterial counts. In contrast, additional administration of antibiotic dramatically decreased blood bacterial counts, but plasma endotoxin levels were concomitantly increased in these animals.

*5.3. LBP Blockade Is a Novel Strategy to Reduce LPS Induced Inflammatory Response.* Our previous study indicated that G-CSF pretreatment induced upregulation of LPS binding protein (LBP) [27]. LPS binding protein (LBP) is an acute phase plasma protein with a molecular weight of 60 KD that can be detected in the acute phase serum of different species such as mice, rabbits, and human [42]. The serum LBP binds to the lipid A component of bacterial endotoxin and facilitates its transfer to the CD14 antigen, which is needed for triggering the inflammatory response via TLR4-NF $\kappa$ B signaling pathway [43]. Martin et al. reported that the binding between LBP and LPS increased the bioactivity of LPS by 100 to 1000 times [44], playing an important role in triggering the inflammatory response in adult respiratory distress

syndrome patients and rabbit, respectively [45]. However, upregulation of LBP, in turn, did augment the inflammatory response [46, 47]. It has been shown in different models that upregulation of LBP prior to an LPS challenge potentiates the inflammatory response which may largely contribute to LPS toxicity in sepsis [46].

Minimizing the inflammatory response to LPS can be achieved by interfering with the interaction of LPS and LBP. The inflammatory response to endotoxin can be decreased by reducing circulating endotoxin levels or by reducing the response of the organism to circulating endotoxin. Experimental strategies include using LBP inhibitory peptide [35], or LPS analogues [48], or using LBP deficient mice [47, 49]. Knapp et al. reported [50] that LBP(−/−) mice were associated with diminished early tumor necrosis factor alpha, interleukin-6, cytokine-induced neutrophil chemoattractant, and macrophage inflammatory protein production and attenuated recruitment of polymorphonuclear leukocytes to the site of infection, indicating that acute inflammation was promoted by LBP. However, LBP(−/−) mice were highly susceptible to *E. coli* peritonitis, as indicated by increased mortality, earlier bacterial dissemination to the blood, impaired bacterial clearance in the peritoneal cavity, and more severe remote organ damage.

Le Roy et al. demonstrated that neutralization of LBP accomplished by blocking either the binding of LPS to LBP or the binding of LPS/LBP complexes to CD14 protected the host from LPS induced toxicity [51]. Araña et al. showed that application of the LBP inhibitory peptides blocked the LBP-LPS interaction efficiently and prevented death of animals in an endotoxin shock model by suppressing the TNF- $\alpha$ response to an LPS challenge [35]. This was confirmed in another publication using CLP-19, a synthetic peptide derived from *Limulus* (anti-LPS factor) [52].

## **6. Outcome of Sepsis Was Improved by Modulating Neutrophil Function and Inflammatory Response**

The role of G-CSF pretreatment and subsequent LBP upregulation was investigated in a SIRS model and a sepsis model. Interestingly, In the SIRS model, G-CSF pretreatment enhanced and accelerated the uptake of LPS by the liver. Subsequently, G-CSF pretreatment caused an overwhelming inflammatory response to LPS leading to the death of all animals in response to an otherwise sublethal dose of LPS. This response could at least be partially attributed to the upregulation of LBP prior to the LPS challenge, as blocking of LBP using an inhibitory peptide abrogated the effects of G-CSF pretreatment [27].

In contrast, in the sepsis model, G-CSF pretreatment was associated with an increased survival rate when compared with an untreated control group [53]. This was paralleled by a reduced inflammatory response. Of note, this response could also be at least partially attributed to the upregulation of LBP prior to the septic insult, as blocking of LBP using an inhibitory peptide abrogated the effect of G-CSF pretreatment. Moreover, LBP upregulation during the infection



FIGURE 2: Balanced modulation of LBP-effect by LBP blockade and G-CSF pretreatment in sepsis. (a) on one hand, LBP promotes bacterial clearance, but on the other hand, it contributes to the sensitization of inflammatory response via activation of NF- $\kappa$ B signaling pathway. (b) Interfering with the LBP-mediated inflammatory response by LBP blockade reduces the inflammatory injury and improves outcome after septic insult.

has seemingly a dual function. Yang et al. found that LBP deficient mice showed delayed neutrophil influx in case of a peritoneal infection [54]. This led to the idea that LBP might have a dual role: augmenting the inflammatory response to bacterial toxin such as LPS and contributing to bacterial elimination via the associated and enhanced neutrophil infiltration (Figure 2). These two functions might help to explain that either LBP blockade or G-CSF treatment alone was not useful in the therapy of patients with severe sepsis.

### **7. Conclusion**

The seeming contradictory results above support the idea that augmenting the neutrophil response via G-CSF treatment is a two-edged sword. Elimination of bacteria by increasing the innate immunity seemed improved via the mobilization of neutrophils, whereas LPS response was augmented via upregulation of LBP [47]. Therefore, these evidences

suggest that the therapeutic strategies by combining the increased bacterial elimination via improving the innate immunity and decreasing the inflammatory injury could be a worthwhile therapeutic strategy for sepsis. Further G-CSFbased therapeutic strategies should be designed to potentially combine with effects of increasing bacterial elimination via mobilization of neutrophil and decreasing the inflammatory response by blocking the LPS response.

#### **Conflict of Interests**

The authors declare that there is no conflict of interests.

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#### **References**

- [1] R. S. Hotchkiss and I. E. Karl, "The pathophysiology and treatment of sepsis," *The New England Journal of Medicine*, vol. 348, no. 2, pp. 138–150, 2003.
- [2] J. Cohen, J. L. Vincent, N. K. Adhikari et al., "Sepsis: a roadmap for future research," *The Lancet Infectious Diseases*, vol. 15, no. 5, pp. 581–614, 2015.
- [3] A. C. Ma and P. Kubes, "Platelets, neutrophils, and neutrophil extracellular traps (NETs) in sepsis," *Journal of Thrombosis and Haemostasis*, vol. 6, no. 3, pp. 415–420, 2008.
- [4] K. Brown, S. Brain, J. Pearson, J. Edgeworth, S. Lewis, and D. Treacher, "Neutrophils in development of multiple organ failure in sepsis," *The Lancet*, vol. 368, no. 9530, pp. 157–169, 2006.
- [5] G. Singer, K. Y. Stokes, and D. Neil Granger, "Reactive oxygen and nitrogen species in sepsis-induced hepatic microvascular dysfunction," *Inflammation Research*, vol. 62, no. 2, pp. 155–164, 2013.
- [6] A. Cauwels and P. Brouckaert, "Nitrite regulation of shock," *Cardiovascular Research*, vol. 89, no. 3, pp. 553–559, 2011.
- [7] M. Aritomi, N. Kunishima, T. Okamoto, R. Kuroki, Y. Ota, and K. Morikawa, "Atomic structure of the GCSF-receptor complex showing a new cytokine-receptor recognition scheme," *Nature*, vol. 401, no. 6754, pp. 713–717, 1999.
- [8] K. Ishikawa, H. Tanaka, Y. Nakamori et al., "Difference in the responses after administration of granulocyte colony-stimulating factor in septic patients with relative neutropenia," *Journal of Trauma-Injury Infection & Critical Care*, vol. 48, no. 5, pp. 814–825, 2000.
- [9] G. S. Chatta, T. H. Price, R. C. Allen, and D. C. Dale, "Effects of in vivo recombinant methionyl human granulocyte colonystimulating factor on the neutrophil response and peripheral blood colony-forming cells in healthy young and elderly adult volunteers," *Blood*, vol. 84, no. 9, pp. 2923–2929, 1994.
- [10] V. Garg, H. Garg, A. Khan et al., "Granulocyte colony-stimulating factor mobilizes CD34<sup>+</sup> cells and improves survival of patients with acute-on-chronic liver failure," *Gastroenterology*, vol. 142, no. 3, pp. 505–512.e1, 2012.
- [11] N. J. Taylor, A. Nishtala, G. K. Manakkat Vijay et al., "Circulating neutrophil dysfunction in acute liver failure," *Hepatology*, vol. 57, no. 3, pp. 1142–1152, 2013.
- [12] M. A. S. Moore, "The clinical use of colony stimulating factors," *Annual Review of Immunology*, vol. 9, no. 1, pp. 159–191, 1991.
- [13] R. Lundblad, J. M. Nesland, and K.-E. Giercksky, "Granulocyte colony-stimulating factor improves survival rate and reduces concentrations of bacteria, endotoxin, tumor necrosis factor, and endothelin-1 in fulminant intra-abdominal sepsis in rats," *Critical Care Medicine*, vol. 24, no. 5, pp. 820–826, 1996.
- [14] P. Villa, C. L. Shaklee, C. Meazza, D. Agnello, P. Ghezzi, and G. Senaldi, "Granulocyte colony-stimulating factor and antibiotics in the prophylaxis of a murine model of polymicrobial peritonitis and sepsis," *Journal of Infectious Diseases*, vol. 178, no. 2, pp. 471–477, 1998.
- [15] A. Bauhofer, A. Torossian, W. Lorenz et al., "Dependence of positive effects of granulocyte colony-stimulating factor on the antibiotic regimen: evaluation in rats with polymicrobial peritonitis," *World Journal of Surgery*, vol. 28, no. 8, pp. 834–844, 2004.
- [16] A. Bauhofer, I. Celik, U. Plaul, H. Wulf, and A. Torossian, "Effects of G-CSF and antibiotic prophylaxis in a  $2 \times 2$  factorial design on outcome in septic rats," *Inflammation Research*, vol. 53, supplement 2, pp. S126–S129, 2004.
- [17] L. Luo, S. Zhang, Y. Wang et al., "Proinflammatory role of neutrophil extracellular traps in abdominal sepsis," *American Journal of Physiology—Lung Cellular and Molecular Physiology*, vol. 307, no. 7, pp. L586–L596, 2014.
- [18] R. K. Root and D. C. Dale, "Granulocyte colony-stimulating factor and granulocyte-macrophage colony-stimulating factor: comparisons and potential for use in the treatment of infections in nonneutropenic patients," *Journal of Infectious Diseases*, vol. 179, supplement 2, pp. S342–352, 1999.
- [19] A. Bauhofer, U. Plaul, A. Torossian et al., "Perioperative prophylaxis with granulocyte colony-stimulating factor (G-CSF) in high-risk colorectal cancer patients for an improved recovery: a randomized, controlled trial," *Surgery*, vol. 141, no. 4, pp. 501– 510, 2007.
- [20] M. V. Relling, J. M. Boyett, J. G. Blanco et al., "Granulocyte colony-stimulating factor and the risk of secondary myeloid malignancy after etoposide treatment," *Blood*, vol. 101, no. 10, pp. 3862–3867, 2003.
- [21] L. Bo, F. Wang, J. Zhu, J. Li, and X. Deng, "Granulocyte-colony stimulating factor (G-CSF) and granulocyte-macrophage colony stimulating factor (GM-CSF) for sepsis: a metaanalysis," *Critical Care*, vol. 15, no. 1, article R58, 2011.
- [22] S. Nelson, S. M. Belknap, R. W. Carlson et al., "A randomized controlled trial of filgrastim as an adjunct to antibiotics for treatment of hospitalized patients with community-acquired pneumonia," *Journal of Infectious Diseases*, vol. 178, no. 4, pp. 1075–1080, 1998.
- [23] R. K. Root, R. F. Lodato, W. Patrick et al., "Multicenter, doubleblind, placebo-controlled study of the use of filgrastim in patients hospitalized with pneumonia and severe sepsis," *Critical Care Medicine*, vol. 31, no. 2, pp. 367–373, 2003.
- [24] P. Hartmann, J. Lammertink, G. Mansmann et al., "A randomized, placebo-controlled study of the use of filgrastim in non neutropenic patients with nosocomial pneumonia," *European Journal of Medical Research*, vol. 10, no. 1, pp. 29–35, 2005.
- [25] A. C. Cheng, D. Limmathurotsakul, W. Chierakul et al., "A randomized controlled trial of granulocyte colony-stimulating factor for the treatment of severe sepsis due to melioidosis in Thailand," *Clinical Infectious Diseases*, vol. 45, no. 3, pp. 308– 314, 2007.
- [26] D. P. Stephens, J. H. Thomas, A. Higgins et al., "Randomized, double-blind, placebo-controlled trial of granulocyte colonystimulating factor in patients with septic shock," *Critical Care Medicine*, vol. 36, no. 2, pp. 448–454, 2008.
- [27] H. Fang, A. Liu, J. Sun, A. Kitz, O. Dirsch, and U. Dahmen, "Granulocyte colony stimulating factor induces lipopolysaccharide (LPS) sensitization via upregulation of LPS binding protein in rat," *PLoS ONE*, vol. 8, no. 2, Article ID e56654, 2013.
- [28] S. Hagel and F. Brunkhorst, "Sepsis," *Intensivmedizin und Notfallmedizin*, vol. 48, no. 1, pp. 57–73, 2011.
- [29] R. P. Dellinger, M. M. Levy, A. Rhodes et al., "Surviving sepsis campaign: international guidelines for management of severe sepsis and septic shock, 2012," *Intensive Care Medicine*, vol. 39, no. 2, pp. 165–228, 2013.
- [30] C. Fleischmann, A. Scherag, N. K. Adhikari et al., "Assessment of global incidence and mortality of hospital-treated sepsis current estimates and limitations," *American Journal of Respiratory and Critical Care Medicine*, 2015.
- [31] R. G. Holzheimer, "Antibiotic induced endotoxin release and clinical sepsis: a review," *Journal of Chemotherapy*, vol. 13, supplement 2, pp. 159–172, 2001.
- [32] K. Buttenschoen, P. Radermacher, and H. Bracht, "Endotoxin elimination in sepsis: physiology and therapeutic application," *Langenbeck's Archives of Surgery*, vol. 395, no. 6, pp. 597–605, 2010.
- [33] Y. Hayashi, "Polymyxin B hemoperfusion (PMX-F) for severe sepsis and septic shock," *Nihon Rinsho*, vol. 70, no. 2, pp. 311– 314, 2012.
- [34] H. Huang, Y.-J. Wang, A. P. White et al., "*Salmonella* expressing a T-cell epitope from Sendai virus are able to induce antiinfection immunity," *Journal of Medical Microbiology*, vol. 58, no. 9, pp. 1236–1242, 2009.
- [35] M. J. Araña, M. G. Vallespi, G. Chinea et al., "Inhibition of LPSresponses by synthetic peptides derived from LBP associates with the ability of the peptides to block LBP-LPS interaction," *Journal of Endotoxin Research*, vol. 9, no. 5, pp. 281–291, 2003.
- [36] E. A. Panacek, J. C. Marshall, T. E. Albertson et al., "Efficacy and safety of the monoclonal anti-tumor necrosis factor antibody  $F(ab')$  2 fragment afelimomab in patients with severe sepsis and elevated interleukin-6 levels," *Critical Care Medicine*, vol. 32, no. 11, pp. 2173–2182, 2004.
- [37] T. Roger, C. Froidevaux, D. Le Roy et al., "Protection from lethal Gram-negative bacterial sepsis by targeting Toll-like receptor 4," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 7, pp. 2348–2352, 2009.
- [38] X. Wittebole, D. Castanares-Zapatero, and P. F. Laterre, "Tolllike receptor 4 modulation as a strategy to treat sepsis," *Mediators of Inflammation*, vol. 2010, Article ID 568396, 9 pages, 2010.
- [39] M. Tidswell and S. P. LaRosa, "Toll-like receptor-4 antagonist eritoran tetrasodium for severe sepsis," *Expert Review of Anti-Infective Therapy*, vol. 9, no. 5, pp. 507–520, 2011.
- [40] A. Barochia, S. Solomon, X. Cui, C. Natanson, and P. Q. Eichacker, "Eritoran tetrasodium (E5564) treatment for sepsis: review of preclinical and clinical studies," *Expert Opinion on Drug Metabolism and Toxicology*, vol. 7, no. 4, pp. 479–494, 2011.
- [41] W. J. Christ, O. Asano, A. L. C. Robidoux et al., "E5531, a pure endotoxin antagonist of high potency," *Science*, vol. 268, no. 5207, pp. 80–83, 1995.
- [42] P. S. Tobias, K. Soldau, and R. J. Ulevitch, "Identification of a lipid A binding site in the acute phase reactant lipopolysaccharide binding protein," *The Journal of Biological Chemistry*, vol. 264, no. 18, pp. 10867–10871, 1989.
- [43] S. M. Opal, J. E. Palardy, M. N. Marra, C. J. Fisher Jr., B. M. McKelligon, and R. W. Scott, "Relative concentrations of endotoxinbinding proteins in body fluids during infection," *The Lancet*, vol. 344, no. 8920, pp. 429–431, 1994.
- [44] T. R. Martin, J. C. Mathison, P. S. Tobias et al., "Lipopolysaccharide binding protein enhances the responsiveness of alveolar macrophages to bacterial lipopolysaccharide. Implications for cytokine production in normal and injured lungs," *The Journal of Clinical Investigation*, vol. 90, no. 6, pp. 2209–2219, 1992.
- [45] J. Fierer, M. A. Swancutt, D. Heumann, and D. Golenbock, "The role of lipopolysaccharide binding protein in resistance to *Salmonella* infections in mice," *The Journal of Immunology*, vol. 168, no. 12, pp. 6396–6403, 2002.
- [46] R. M. Minter, X. Bi, G. Ben-Josef et al., "LPS-binding protein mediates LPS-induced liver injury and mortality in the setting of biliary obstruction," *The American Journal of Physiology— Gastrointestinal and Liver Physiology*, vol. 296, no. 1, pp. G45– G54, 2009.
- [47] R. S. Jack, X. Fan, M. Bernheiden et al., "Lipopolysaccharidebinding protein is required to combat a murine gram-negative bacterial infection," *Nature*, vol. 389, no. 6652, pp. 742–745, 1997.
- [48] J. Hellman and H. S. Warren, "Antiendotoxin strategies," *Infectious Disease Clinics of North America*, vol. 13, no. 2, pp. 371–386, 1999.
- [49] F. Pène, E. Courtine, F. Ouaaz et al., "Toll-like receptors 2 and 4 contribute to sepsis-induced depletion of spleen dendritic cells," *Infection and Immunity*, vol. 77, no. 12, pp. 5651–5658, 2009.
- [50] S. Knapp, A. F. de Vos, S. Florquin, D. T. Golenbock, and T. van Der Poll, "Lipopolysaccharide binding protein is an essential component of the innate immune response to *Escherichia coli* peritonitis in mice," *Infection and Immunity*, vol. 71, no. 12, pp. 6747–6753, 2003.
- [51] D. Le Roy, F. Di Padova, R. Tees et al., "Monoclonal antibodies to murine lipopolysaccharide (LPS)-binding protein (LBP) protect mice from lethal endotoxemia by blocking either the binding of LPS to LBP or the presentation of LPS/LBP complexes to CD14," *The Journal of Immunology*, vol. 162, no. 12, pp. 7454–7460, 1999.
- [52] Y. Liu, B. Ni, J.-D. Ren et al., "Cyclic limulus anti-lipopolysaccharide (LPS) factor-derived peptide CLP-19 antagonizes LPS function by blocking binding to LPS binding protein," *Biological and Pharmaceutical Bulletin*, vol. 34, no. 11, pp. 1678– 1683, 2011.
- [53] A. Liu, S. Weiss, H. Fang et al., "Lipopolysaccharide-binding protein (LBP) blockade augments the protective effect of granulocyte colony-stimulating factor (G-CSF) in a rat sepsis model," *Shock*, vol. 43, no. 5, pp. 497–503, 2015.
- [54] K. K. Yang, B. G. Dorner, U. Merkel et al., "Neutrophil influx in response to a peritoneal infection with Salmonella is delayed in lipopolysaccharide-binding protein or CD14-deficient mice," *Journal of Immunology*, vol. 169, no. 8, pp. 4475–4480, 2002.
- [55] T. Hartung, S. Von Aulock, C. Schneider, and E. Faist, "How to leverage an endogenous immune defense mechanism: the example of granulocyte colony-stimulating factor," *Critical Care Medicine*, vol. 31, no. 1, supplement, pp. S65–S75, 2003.
- [56] H. Tanaka, M. Nishino, Y. Nakamori et al., "Granulocyte colonystimulating factor (G-CSF) stiffens leukocytes but attenuates inflammatory response without lung injury in septic patients," *Journal of Trauma-Injury Infection & Critical Care*, vol. 51, no. 6, pp. 1110–1116, 2001.



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